

Insights into the functional role of tea microbes on tea growth, quality and resistance against pests and diseases

Xiaoyang ZHANG^{1,2}, Haozhi LONG^{1*}, Da HUO^{1,3}, Masood I. AWAN⁴,
Jinhua SHAO^{1,5}, Athar MAHMOOD⁶, Shuang LIU², Jigang HUANG²,
Alia PARVEEN⁴, Muhammad AAMER⁷, Muhammad U. HASSAN⁷

¹Jiangxi Agricultural University, Nanchang 330045, China; zxygo123@163.com;

longhaozhi1987@126.com (*corresponding author); bawda1090@163.com; sbaojinhua@163.com

²Jiujiang Agricultural Technology Extension Centre, Jiujiang 332000, China; jjzbzj@163.com; zxygogo123@126.com

³College of Tea and Food Science (Wuyi University), Nanping 354300, China

⁴University of Agriculture Faisalabad, Department of Agronomy, Sub-Campus Depalpur, Okara, Faisalabad, Pakistan;

Masood.awan@uaf.edu.pk; aliachaudhry155@gmail.com

⁵China Guangxi Hydraulic Research Institute, Key Laboratory of Water Engineering Materials and Structures, Nanning, 530023, China

⁶University of Agriculture Faisalabad, Department of Agronomy, Faisalabad, Pakistan; athar.mahmood@uaf.edu.pk

⁷Jiangxi Agriculture University, Research Center on Ecological Sciences, Nanchang 330045, China; muhammadaamer@jxau.edu.cn;

muhassanuaf@gmail.com

Abstract

Tea is an economical and most widely used beverage across the globe owing to its unique fragrance and flavor. Plant microbe interaction has emerged as an important topic which got the attention of scientists to improve plant performance. Tea microbes remained a prominent research topic for scientists over the years as tea microbes helps in nutrient cycling and stress management which in turn improve the tea growth, yield and quality. The roots of tea plants are colonized by various microbes including arbuscular mycorrhizal fungi (AMF), bacterial communities, and endophytes increase root growth, development and nutrient uptake which in turn improve tea growth, yield and quality. These microbes also increase the concentration of nutrients, amino acids, soluble proteins, flavonoids, catechuic acid, glucose, fructose, sucrose contents caffeine, and polyphenols concentration in tea plants. Besides this, these microbes also protect the tea plants from harmful pest and diseases which in turn leads to an appreciable improvement in plant growth and development. The most important goal of any farming system is to establish a system with production of maximum food while minimizing impacts on the environment. The present review article highlights the role of various microbes in improving the growth, yield and quality of tea plants. In addition, we also discussed the research gaps to improve our understanding about the role of tea microbes in improving tea growth, yield, pest and diseases resistance. We believe that this review will provide a better insight into the existing knowledge of tea microbes in improving tea growth and yield.

Keywords: AMF; disease; growth; microbes; quality; tea

Received: 14 Sep 2022. Received in revised form: 24 Oct 2022. Accepted: 31 Oct 2022. Published online: 05 Dec 2022.

From Volume 49, Issue 1, 2021, Notulae Botanicae Horti Agrobotanici Cluj-Napoca journal uses article numbers in place of the traditional method of continuous pagination through the volume. The journal will continue to appear quarterly, as before, with four annual numbers.

Introduction

Tea is an important and extensively cultivated beverage across the globe (Bora and Bora, 2021). It is an imperious economic crop widely grown in China and South Asia and it contains a significant amount of phenolic compounds and minerals (Wan, 2003; Chen and Lin, 2015). Tea is mainly produced in Asia, Africa, Caspian Seas and South American countries. China, India, Kenya and Sri Lanka are the major tea producing countries and they have a share of 75% in global tea production. Globally, 6,497,443 tons of tea are produced every year which has a global market value of 207.1 billion USD (FAO, 2020). Tea is long duration crop and it is subjected to diverse pests and disease attack which cause a substantial reduction in crop growth and yield (Mareeswaran *et al.*, 2015). Globally, inorganic chemicals have been used to remedy this problem (Adesoye and Kloepper, 2009). However, the application of chemicals to tea is prohibited owing to various reasons including soil deterioration, ground water pollution, residual effects in tea plants followed by the development of resistance and outbreak of secondary pests (Gurusubramanian *et al.*, 2005; Hazarika *et al.*, 2009; Saha and Mukhopadhyay, 2013). Moreover, the inappropriate use of chemicals to control pests and diseases also poses a serious threat to soil health and the environment. Therefore, now focus has changed to improve crop performance and manage diseases by using economical and ecologically safe methods (Bag *et al.*, 2022).

Considerable efforts are being made across the globe through the use of alternative biological approaches, agricultural practices and microbes to improve crop growth, maintain soil health and productivity and reduce the infestation of pests and diseases (Fernando *et al.*, 2005). Soil is a dynamic natural environment having an intense plant-microbes interaction that significantly affects the crops by initiating the production of growth hormones and harnessing essential nutrients (Bhattacharyya and Jha, 2012). Different microbes including rhizobia, arbuscular mycorrhizal fungi (AMF), actinomycetes and diazotrophic bacteria play an imperative role in nutrient cycling and their availability to host which in turn improves plant growth (Saharan and Nehra, 2011). Besides this soil microbes also affect the soil physio-chemical and biological properties which positively affect the plant growth and development (Saharan and Nehra, 2011).

Tea rhizo-sphere has been investigated recently in terms of plant microbe interactions. The growth of tea is hampered by soil temperature, poor nutrient availability and pruning. It has been documented that AMF significantly improve plant growth by increasing the plant nutrient uptake capacity (Shao *et al.*, 2021) and nutrient availability to plants (Bag *et al.*, 2022). AMF association significantly improves the nutrient mobilization and ensures better nutrient availability thus resulting in a significant increase in plant growth and development (Abbasi *et al.*, 2015). Recently, endophytes also emerged as important microbes and it has been noted that colonize plants and produce secondary metabolites microbiologists and also provide protection to plants against different pests and diseases (Yan *et al.*, 2018). Endophytes also improve the plant growth by nitrogen fixation, phosphorus solubilization, and production of indole-3-acetic acid (IAA), and improving the activity of 1-aminocyclopropane-1-carboxylate (ACC) deaminase which in turn improve the plant growth and development (Yan *et al.*, 2018).

The rhizosphere is considered to be the heart of soil and it has direct plant microbe interaction. The plant roots provide photosynthates for microbial growth which in return facilitate plants growth and subsequent development (Banik *et al.*, 2014; Olanrewaju *et al.*, 2019). Besides this, microbes also play an important role as biocontrol agents to control various pathogens and diseases (Rungsirivanich *et al.*, 2020). The identification of variety as well structural make of microbes is critical for ensuring their long-term use in tea gardens. The present review highlights the importance of various known and putative microbes native to tea soils for improving the growth, yield and quality of tea and resistance to pests and diseases. Moreover, we also shed light on various research gaps that must be fulfilled in future research studies for improving tea growth, yield and quality.

The role of soil microbial inoculants as bio-fertilizers

Soil is a rich environment and its health is significantly affected by different parameters including microbial growth, root exudates and different abiotic and as well as biotic factors (Bhattacharyya and Jha, 2012). The diversity of microbes and microbial interactions in soil are very complex and they significantly affect crop growth and soil health (Bhattacharyya *et al.*, 2014). The microbes are considered essential for any ecosystem owing to role in biogeochemical cycles, nutrient acquisition, decomposition of soil organic matter (SOM), water uptake, and pest and disease control. The microbial bio-pesticides and bio-fertilizers are considered as an effective tool for pest management (Bhardwaj *et al.*, 2014). Microbial bio-fertilizers improve the N fixation, solubilization of phosphorus, production of growth promoting hormones, nutrients and water uptake which in turn improve plant growth and productivity (Miransari, 2011; Dodd and Ruiz-Lozano, 2012).

Microbial bio-fertilizers contain live micro-organisms and their field application improves the plant's growth, soil fertility and plant tolerance ability by increasing the supply of nutrients (Bhardwaj *et al.*, 2014; Vessey, 2003). The microbial bio-fertilizers are considered to be safe because they help to reduce global warming and reduce use of chemicals and prevent the degradation of SOM (Mia *et al.*, 2010). The microbial bio-fertilizers include PGPR (plant growth promoting rhizobacteria), AMF and actinomycetes which play a significant role in nutrient cycling and plant growth and development (Mishra *et al.*, 2013). The application of PGPRs substantially improves the N and phosphorus (P) uptakes which indicate their beneficial impacts on soil nutrient cycling and their availability to plants (Rajendran and Devaraj, 2004). The tea industry has achieved a significant success in exploitation of bio-fertilizers and in recent years the use of bio-fertilizers has substantially increased the tea growth and productivity (Bhardwaj *et al.*, 2014). Various strains of PGPR (*Bacillus pseudomycooides*, *Burkholderia*, *Enterobacter lignolyticus* and *Pseudomonas aeruginosa*) are being used as bio-fertilizers to improve the growth and productivity of tea (Chakraborty *et al.*, 2013; Dutta *et al.*, 2015). Likewise, the use of *Azotobacter chroococcum*, *Bacillus subtilis* and *Pseudomonas* has also increased the growth and production of tea (Nepolean *et al.*, 2012; Pandey *et al.*, 2013).

Tea plants treated with microbes showed a significant increase in plant girth, root length, productivity of young shoots (Pandey *et al.*, 2013). The use of N-fixing bacteria (NFB), P-solubilizing microbes (PSMs), K-solubilizing microbes have a tremendous potential and their inoculation to tea plants significantly improve soil health, microbial growth, plant growth, yield and quality (Baby, 2002; Aggani, 2013). The various methods including seed treatments, soil and foliar sprays are used to apply microbial bio-fertilizers to tea plants. However, the application method depends on different cultural practices and type of plant materials (Mondal *et al.*, 2015; Dutta *et al.*, 2015). SOM plays a significant part in soil health and proliferation of microbes and soil rice in organic matter (OM) are considered to have luxuriant growth of beneficial microbes whereas soil with poor OM status has reduced and restricted growth of microbes (Dutta *et al.*, 2015). In conclusion, soil microbes, improves nutrient availability and soil organic matter which in turn improve the growth and yield of tea.

Role of AMF in tea growth, and quality

It has been reported that 80% of plant species develop AMF association (Wang and Jiang, 2015). Tea plants colonized with AMF have better root growth, nutrient and water uptake resulting in a substantial increase in tea growth and development (Shao *et al.*, 2019). AMF also increased the amino acid contents in tea plant leaves followed by an increase in genes expression involved in amino acid synthesis enzymes. Moreover, AMF also appreciably increases the concentration of total soluble proteins (TSP), glucose, phenolic and flavonoid in the leaves of tea plants (Shao *et al.*, 2019). AMF generally considered as bio-fertilizers and AMF inoculation offers resistance to host plants against different biotic and abiotic stresses (Chandra and Banik,

2021). The significance of AMF in regulating the plant growth and development has gotten significant attention in recent times (Wu *et al.*, 2019).

AMF colonization significantly improves root length, root dry weight and root volume which in turn improves the nutrient and water uptake and resulting in significant increase tea growth and development (Abbasi *et al.*, 2015). AMF increases plant growth and development in normal and stressed conditions. AMF inoculation also reduce the use of chemical fertilizers and improve the soil health and therefore improve the tea growth, yield, leaf nutrient concentration and leaf quality (Bag *et al.*, 2022). AMF colonization also increases the synthesis of flavonoid contents and it also increases the accumulation different secondary metabolites in tea plants (Zubek *et al.*, 2015). The concentration of amino acids is also increased the tea plants inoculated with AMF (Baslam *et al.*, 2013; Sanmartín *et al.*, 2014).

Moreover, concentration of leaf nutrients (Shao *et al.*, 2019) and whole polyphenol contents are also substantially increased in tea plants inoculated with *G. mosseae* (Zhao *et al.*, 2014). Theanine is an imperative metabolite produced by tea plants (Sharma *et al.*, 2018). Many AMF species including *Glomus intraradices*, *Glomus versiforme* and *Glomus etunicatum* substantially improved theanine, copper, iron and zinc contents in tea plant tissues (Dash *et al.*, 2019). AMF positively affect nutrient acquisition and improves water uptake which leads to an appreciable increase in tea growth, yield and quality (Wu *et al.*, 2019). Salt stress causes a substantial reduction growth of tea, nonetheless, AMF significantly improves osmolyte accumulation, and reduces the lipid per-oxidation and improves salinity tolerance (Liu *et al.*, 2014).

AMF considerably increased the P concentration, encourage P absorption which reduce dependence on inorganic fertilizers and leading to significant improvement in soil health, nutrient availability and tea growth and yield (Mei *et al.*, 2019). AMF also improves the tea quality and it also induced a significant impact on growth, yield and tea quality. Recently, it has been reported that AMF improves concentration of phytohormone, nutrient absorption, root growth, genes expression and tea biomass production (Shao *et al.*, 2021). Moreover, AMF also increases concentration of polyphenol, amino acids, flavonoids, catechuic acid, TSP, glucose, fructose and sucrose contents (Abbasi *et al.*, 2015; Wu *et al.*, 2019; Begum *et al.*, 2019; Phour *et al.*, 2020). To summarize, microbes improve nutrient uptake, root growth and osmolytes accumulation thereby improve the tea growth and yield.

Role of bacterial communities in growth and quality of tea

Bacterial communities also play a crucial role in tea growth and yield. The bacterial families like *Pedospaeraceae*, *Solibacteraceae*, and *Gemmataceae* play a significant role in tea growth and development (Bhattacharyya *et al.*, 2020). Recently, it has been reported that bacterial strain (*Serratia marcescens*: ETR17 strain) significantly improved the tea growth and development and resistance against tea root and foliage pests (Dhar *et al.*, 2018). In another study, *Burkholderia pyrrocinia* (P10 strain) obtained from the tea region showed an appreciable potential to be used as bio-fertilizer (Han *et al.*, 2021). Tea rhizosphere has a wide range of bacteria including *Bacillus*, *Beijerinckia*, *Azospirillum*, *Comamonas acidovorans*, *Pseudomonas aeruginosa* and *Paenibacillus polymyxa* which improve the tea growth (Table 1) and development by increasing production of IAA, siderophore, solubilization increasing resistance against pathogens (Chakraborty *et al.*, 2012; Morang *et al.*, 2015).

The bacteria colonizing the tea roots also produce growth promoting substances including terpenoids, steroids and diterpenes which substantially improved the tea growth and yield (Tshikhudo *et al.*, 2019). In a recent study, *Pseudomonas* and *Bacillus* isolated from tea rhizosphere showed a significant potential to improve the plant growth and yield of tea plants (Vandana *et al.*, 2018). The native strains of tea rhizosphere region improved the growth of tea plants. The *Bacillus* species is a viable option to improve tea growth owing to their appreciable ability to produce spores and sensitivity to different climate conditions whereas, *Pseudomonas*

species are considered to be alternate option owing to their better colonization capacity and their ability to suppress the fungal diseases (Vandana *et al.*, 2018; Arafat *et al.*, 2020). In another study, it was noted *Enterobacter lignolyticus* strain collected from tea rhizosphere significantly increased the growth and development of tea clones (Dutta *et al.*, 2015). Likewise, inoculation of tea plants with PGPR significantly improved the growth, yield and quality of tea plants (Chakraborty *et al.*, 2006; Chakraborty *et al.*, 2013). Bacterial strains like, *Brevibacterium* and *Lysinibacillus* have been found to possess important PGRP characteristics and they significantly improve the plant growth by production of IAA and solubilization of P (Borah *et al.*, 2019).

Chopra *et al.* (2020) identified 23 isolates of bacteria from the Assam tea garden. The collected strains were tested for their potential to improve plant growth and their anti-fungal properties against fungal pathogens (*Fomeslomaensis*, *Corticium rolfsii*, and *Rhizoctonia solani*). They noted that *Brevibacterium sediminis* has excellent potential as PGPR and it also possesses excellent antifungal properties (Chopra *et al.*, 2020). In another study, Shan *et al.* (2018) obtained a wide range of actinobacteria from tea plants. They identified 46 different actinomycetes and most common genus was Streptomyces. *Piscicoccus* and *Mobilicoccus*. These authors noted that this strain have higher capacity to produce secondary metabolites and they also significantly increased the IAA production, and possess excellent anti-bacterial and anti-fungal activities (Shan *et al.*, 2018).

Table 1. Effect AMF on growth, yield and quality of tea

AMF Species	Effects	Reference
<i>Claroideoglossum etunicatum</i>	AMF inoculation increased plant height, root and shoot biomass, total leaf area, root volume and leaf N, P, K, Ca, Mg, Mn and Zn contents of tea plants.	(Shao <i>et al.</i> , 2018)
<i>Acaulospora scrobiculata</i> (A.s.) <i>Glomus macrocarpum</i> (G.m.), <i>Rhizophagus intraradices</i> (R.i.)	AMF inoculation improved leaves per plant, leaf area, plant height, shoot and root length, and fresh and dry weight of roots and shoots.	(Sharma and Kayang, 2017)
<i>Clariodeoglossum etunicatum</i>	AMF inoculation in drought stress conditions in tea plant improved antioxidant enzyme (superoxide dismutase (SOD), catalase (CAT), guaiacol peroxidase, and ascorbate peroxidase) activities, AMF enhanced plant defense mechanism which ultimately and reduced the oxidative damage of drought stress	(Chun-Yan <i>et al.</i> , 2020)
<i>Glomus versiforme</i>	AMF inoculation under salinity conditions improved growth, root and leaf N, P, Fe, Zn, and Mg. AMF also reduced water saturation deficit	(Liu <i>et al.</i> , 2013)
<i>Glomus etunicatum</i>	AMF inoculation in tea plant under shading enhanced the plant height, root and shoots biomass, root length and volume, nutrients uptake except Fe, and stress resistance.	Sun <i>et al.</i> , 2020)
<i>Glomus mosseae</i>	AMF inoculation alleviated the salinity stress in tea and improved the quality of tea leaves.	(Guo <i>et al.</i> , 2021)
<i>Glomus epigaeumg</i>	AMF inoculation improved plant biomass, chlorophyll, leaf P, Fe, Zn, Mg and Cu. AMF also enhanced soil phosphatase activity	(He and Liu, 1994)
<i>Glomus mosseae</i> , <i>Glomus intraradices</i>	AMF inoculation increased the callusing rate, chlorophyll content, and root resistant against pathogens	(Chelangat <i>et al.</i> , 2021)
<i>Claroideoglossum etunicatum</i>	AMF showed significant higher growth in root length, root and shoot biomass, plant height. AMF also increase phosphatase activity	(Shao <i>et al.</i> , 2021)

<i>Acaulospora spinosa</i> , <i>Glomus aggregatum</i> , <i>Glomus ambisporum</i> , <i>Glomus clavisporum</i> , <i>Glomus geosporum</i>	AMF in acidic soils increased root/shoot length, dry weight; root /shoot ratio, caffeine contents, sugar contents polyphenols contents which improved the tea quality	(Singh <i>et al.</i> , 2010)
--	---	------------------------------

AMF: arbuscular mycorrhiza fungi, N: nitrogen, P: phosphorus, K: potassium, Ca: calcium, Mg: magnesium, Mn: manganese, Zn: zinc, Fe, iron, Cu: copper.

Colletotrichum also colonize the tea plants and they significantly improved the tea growth, yield and quality (Liu *et al.*, 2015). Likewise, different N-fixing bacteria (*Azospirillum*) and P-solubilizing bacteria were identified from Indian tea soils and they significantly increase plant growth and resistance against pest and diseases (Cernava *et al.*, 2019). In conclusion, bacterial communities improve nutrient uptake, and synthesis of hormones, resulting in substantial improvement in yield and quality.

Role of endophytes in growth and development of tea

Tea plants also host different groups of endophytic microbes (bacteria & fungus) in their leaves, roots and stem. The diversity of endophytes colonizing the tea plants depends on tea genotypes, tissue specification and seasonal variations (Yan *et al.*, 2018). The endophytic bacteria colonizing tea plants can be categorized into seven classes, four phyla, thirteen orders, thirty-two genera and twenty-four families (Xie *et al.*, 2020). However, the most predominant members colonizing the tea are from four different orders including *Bacillales*, *Burkholderiales*, *Micrococcales*, and *Rhizobiales*. Moreover, members from orders of *Glomerellales*, *Xylariales*, and *Diaporthales* are major contributors in the various tissues of tea plants (Xie *et al.*, 2020).

Endophytic microbes provide secondary metabolite to plants and leading to a significant increase in tea growth (Rustamova *et al.*, 2020). Besides this, they also generate different growth regulators including siderophores, ammonia, ACC deaminase and phytohormones and improve the efficiency of macro and micro-nutrients which in turn improve the plant growth and development (Alurappa *et al.*, 2018; Banik *et al.*, 2019a). Endophytic fungus colonizing the tea plants also produces secondary metabolites which improve the plant growth and flavor of tea plants (Boruta, 2018; Xie *et al.*, 2020). The microbial mediated increase in phytohormones is a promising option to improve the tea growth and yield (Figure 1) and tea endophytes is a viable option to induce the production of phyto-hormones which in turn improve tea growth and production (Mu *et al.*, 2021). Moreover, tea endophytes also produce volatile compounds with a biological activity which has tremendous potential to be used against different pathogenic agents including bacteria, fungus and pests (Mu *et al.*, 2021). The endophytic fungi colonizing the tea plants improve the tea growth and productivity and this endophytic fungus also potential to be used as bio-fertilizers for other crops (Nath *et al.*, 2015). In crux, endophytes improve the synthesis of siderophores, ammonia, ACC deaminase and phytohormones and the efficiency of macro and micro nutrients thus leading to a significant increase in growth and yield.

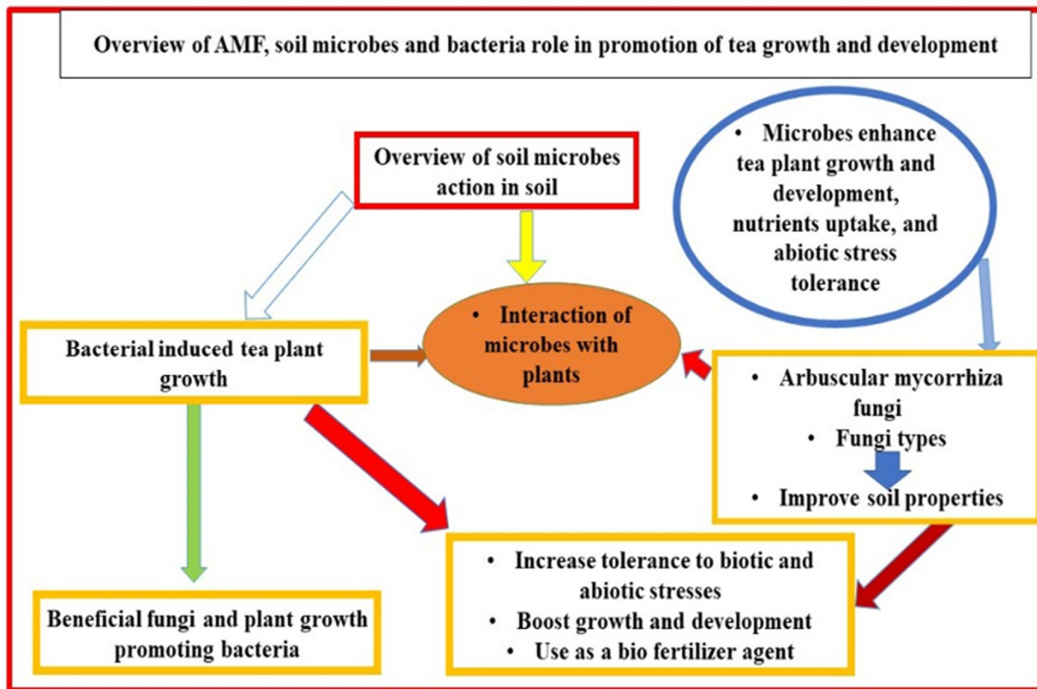


Figure 1. The soil microbes including AMF and bacteria improves, nutrient uptake, soil properties and water uptake which in turn improve the growth and stress tolerance in tea

Role of tea microbial pesticides to control tea-associated pests

Tea is long duration crop which is subjected to different pests that cause huge yield losses (Banik *et al.*, 2019b). The insects and pests pose a serious threat to tea cultivation that causes huge yield losses. Mites and insect pests affect tea cultivation and they can cause yield losses from 5-55% (Idris *et al.*, 2020). Microbial pesticides increase growth of plants by suppressing the activities of tea pests and pathogenic agents (Bhattacharyya and Sarmah, 2018). Mites and pests cause damaging effects to tea plants (Yu *et al.*, 2018) and *Ectropis obliqua* and *Gyropsylla spegazziniana* are prime mite and pests affecting the tea plants (Kakoki *et al.*, 2018). The microbes were discovered as an important biological weapon to control the pests of tea around 1970. The microbial pesticides are mostly produced by different entomopathogenic microbes (Idris *et al.*, 2020) and more than 40 bacterial and fungal taxa and 82 viral species efficient against tea pests have been identified (Idris *et al.*, 2020). The microbial agents including *Bacillus thuringiensis*, *Paecilomyces Beauveria*, *Euproctis* and *pseudoconspira* are important microbes to control the pests of tea plants (Banik *et al.*, 2014). The sustainable cultivation of tea focuses on the use of environment friendly approaches to control the pests of tea and reduce reliance on the use of synthetic pesticides.

Table 2. Effect of different microbes on growth, yield and quality of tea

Microbial species	Effects	Reference
Bacteria (BF9: <i>Bacillus megaterium</i> 47/9+ <i>Paenibacillus macquariensis</i> RC696)	Bacterial strains stimulated tea plant growth, leaf area, green leaf yield, chlorophyll contents and enzyme activities.	(Çakmakci <i>et al.</i> , 2021)
Basidiomycetes and actinomycetes	Interact with each other and improve the quality of tea	(Wang, 2019)
Bacteria (Azoto II-1, <i>Acinetobacter</i> sp., <i>bacteria</i> Endo-5, <i>bacteria</i> Endo-65 and Endo-76)	Bacteria improved the shoot growth of tea up to 17.26% higher than the control.	(Fauziah <i>et al.</i> , 2019)
<i>Cyberlindnera</i> , and <i>Aspergillus</i> , <i>Uwebraunia</i>	Increased activities of polyphenol oxidase, cellulase, and pectinase as promoted by bacteria genera	(Li <i>et al.</i> , 2018)
<i>Pseudomonas fluorescens</i>	Bacteria produce antibiotics, chelation of available iron, and synthesis of extracellular enzymes	(Bhattacharyya and Jha, 2012)
<i>Okons</i> (<i>Azospirillum</i>)	Enhance the nutrient level in tea seedlings or cuttings which resulted in higher yield and quality	(Phukan <i>et al.</i> , 2012)
<i>Bacillus</i> RC23 and <i>Paenibacillus polymyxa</i> RC05	Microbial inoculation improved the root length, number of roots, root quality, rooting percentage and root dry weight.	(Erturk <i>et al.</i> , 2008)
<i>Paenibacillus</i> , and <i>Arthrobacter</i>	These bacterial types showed high variability in utilizing various carbon sources in tea which increase yield.	(Çakmakçı <i>et al.</i> , 2010)

The use of microbes is a fascinating approach to control the pests of tea and in recent times it got attention around the globe (Bhattacharyya and Sarmah, 2018). Diverse microbial control methods have been acknowledged in various tea growing systems to produce pesticide free tea (Bhattacharyya and Sarmah, 2018). Moreover, to a great extent, production and utilization of microbial combinations is a beneficial option to improve tea growth and resistance against different pests (Bhattacharyya and Sarmah, 2018).

Role of tea microbial pesticides to control tea-associated diseases

Diseases are important biotic stresses that are causing huge growth and yield losses to tea crops across the globe. The intensity and incidence of diseases vary with climatic conditions, elevation and planting materials. Tea is susceptible to different tea pathogens and microbial tea rhizosphere also has many PGPR that improves the tea growth (Table 2) and resistance against different tea diseases (Phukan *et al.*, 2012). Various PGPR including *Aspergillus*, *Azotobacter*, *Azospirillum*, *Fusarium*, *Gliocladium Penicillium*, *Trichoderma* and *Pseudomonas* have appreciable potential to improve tea growth and resistance against tea pathogens (Mokhtar and El-Mougy, 2014). For instance, *Bacillus subtilis* significantly improves the resistance of tea against different diseases of tea including black rot, blister blight, branch canker, thorny stem blight and root diseases (Barthakur, 2011).

Sowndhararajan *et al.* (2013) studied the use of different biocontrol agents to control the diseases of tea. They used *Ochrobactrum anthropi* in combination with chemical fungicides to control the blister blight disease in tea. These authors noted that *Ochrobactrum anthropi* significantly managed the tea blister blight disease of tea. Likewise, Dutta *et al.* (2012) also found that the use of *Pseudomonas aeruginosa* managed brown rot disease of tea and improve resistance of tea plants against this disease (Dutta *et al.*, 2012). Anita *et al.* (2012) also documented that *T. atroviride* produce the secondary metabolites and enzymes that induce the resistance

against the Phomopsis canker (Anita *et al.*, 2012). The synthesis of various enzymes including amylase, cellulase, chitinase, polygalacturonase and protease is linked with the antagonist activities of bio-control agents. For instance, *Bacillus* and *Pseudomonas* improved the resistance of tea plants against tea blight by increasing the synthesis of the aforementioned enzymes (Pallavi *et al.*, 2012).

Saravanakumar *et al.* (2007) tested the efficiency of *Pseudomonas* and *Bacillus* against blister blight disease of tea. They noted that these microbes significantly reduce the severity of blister blight which was linked with an increase in the synthesis of chitinase, glucanase, peroxidase (POD), phenolics and phenylalanine ammonia lyase (PAL). In another study, Premkumar *et al.* (2012) studied the efficacy of *Bacillus*, *Pseudomonas* and *Trichoderma* against the different fungal pathogens. The use of these microbes significantly increased the resistance and reduced the severity of eye spot disease of tea plants (Gnanamangai and Ponmurugan, 2012). Actinomycetes also have appreciable potential as bio-control agent and they can significantly manage the disease of tea (Bhattacharyya *et al.*, 2015). The *actinomycetes* including *Streptomyces nojiriensis*, *Streptomyces griseoluteus* and *Streptomyces somaliensis* are very effective (Figure 2) to control the different diseases of tea (Barthakur, 2011). The use of microbes to control diseases is an effective approach and it reduces dependence on chemical substances and it also improves the quality of soil and quality of tea production. The application of microbes is crucial to soil type and soil type significantly affects the efficiency of microbes to suppress diseases (Wardle *et al.*, 2004; Hazarika *et al.*, 2009). In the light of aforementioned findings increases the synthesis of different enzymes and nutrient availability and uptake which in turn improve tea growth and yield.

Interaction between tea rhizosphere microorganisms and environmental factors

Soil nutrients, pH, and water holding capacity are important components of soil that significant affect the abundance of soil microbes (Greenlon *et al.*, 2019). The microbe's diversity in soil rhizosphere has strong association with environmental factors (Figure 3). The soil properties are significantly changed owing to different rhizobacteria and the application of chemical fertilizers (Wang *et al.*, 2021). Soil microbes play an appreciable role in ecosystem stability, soil heath and soil functioning. The planting species, soil properties and farming practices significantly affect the microbial population in tea rhizosphere. Different authors noted a complex relationship between tea microbes in environmental factors in tea plantations (Li *et al.*, 2016). Recently, the rhizosphere of tea has been studied in detail and noticeably from the prospective of plant microbe interactions. These discoveries have opened a new window for future research in the domain of soil microbe interaction in tea plantations. The abundance and variety of microbes colonizing the tea plants is varied owing to geographical surroundings (Xie *et al.*, 2020).

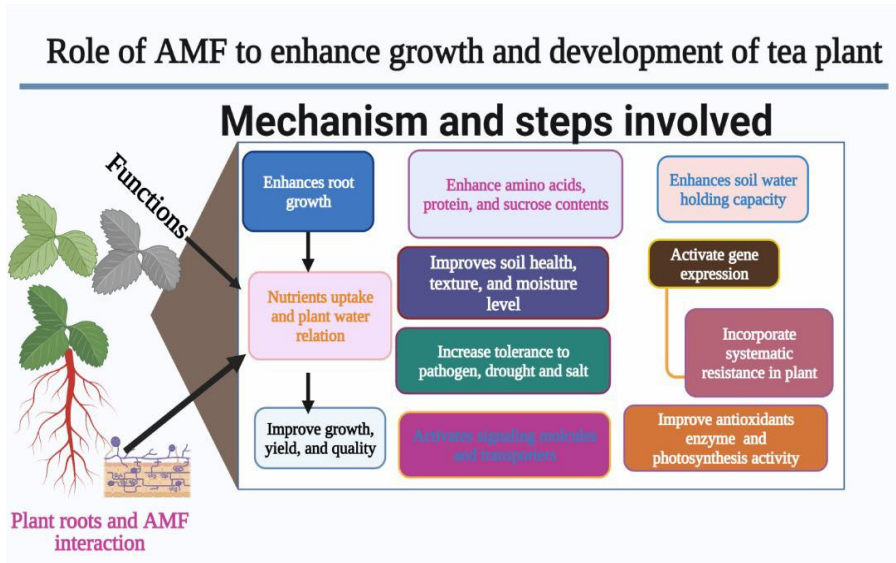


Figure 2. AMF enhances root growth, nutrient uptake, soil water holding capacity, genes expression, photosynthetic efficiency and antioxidant activity therefore improve the tea growth, yield, and quality and resistance against stresses

The long-term tea production boosted the abundance of microbial N and carbon while decreases the metabolic quotient (Li *et al.*, 2017). The most abundant bacteria genera found in tea plantations are *Bacillus*, *Paenibacillus*, *Enterobacter*, *Pseudomonas* and *Acinetobacte* and they effectively improved the soil nutrient status, growth and yield of tea plants (Shang and Liu, 2021). PGPRs not only improve the soil nutritional status but they also restrict the growth of tea infections by producing various antibiotics, antifungal and insecticide compounds (Wang *et al.*, 2021). Recently, it has been documented that degraded soil ecosystems can be remediated by using microbial abundance (Wang *et al.*, 2019). Thus, the addition of organic matter, microbial fertilizers and the selection of suitable varieties can alleviate the issues of tea plantations (Bag *et al.*, 2022).

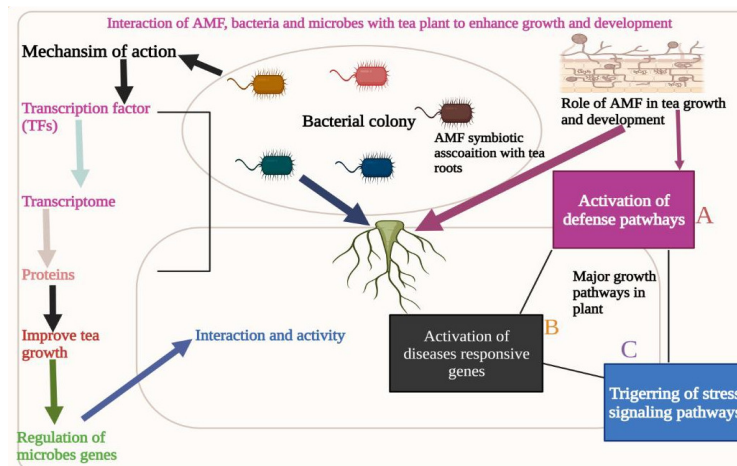


Figure 3. Interaction of AMF, bacteria and microbes in tea rhizosphere. Microbes enter into cell of tea plants by molecular receptors and they modulate genes expression pathways that are linked with defense responses, resistance against diseases and growth promotion of tea plants. The microbes also modulate the genes expression which influences the environment as well as growth of plants

Use of microbe's diversity sustainable production of tea

Microorganisms play a critical role in the agricultural production system. The development of a balanced and productive ecosystem is crucial for the survival of earth. The world is transferring to ecologically safe and economical production systems to increase the per unit production of tea (Gebrewold, 2018). The extensive use of chemicals induces negative impacts of soil health and their prolonged use also impose negative impacts on soil nutrient status and microbial diversity. Therefore, in this context use of modern production practices and biotechnological approaches are in high demand to improve soil health, growth and yield of crops. A huge quantity of synthetic fertilizers has been used over the years for the plantation of tea and long-term use of chemical substances has degraded the soil quality and tea yield (Chakraborty *et al.*, 2013). Therefore, there is dire need to reduce the use of chemical substances in tea production and replace it biological agents to enhance soil fertility, tea quality and resistance against pests and diseases (Vandana *et al.*, 2018).

AMF, microbes and PGPR showed beneficial impacts on tea growth, soil health and the environment. These aforementioned microbes also enhanced the immunity of plants by affecting the diverse cellular metabolisms and preventing the attack of pests and diseases. PGPR are most effective and promising microbes that improve the plant growth and soil without contaminating our environment region (Vandana *et al.*, 2018). The increase in plant growth and resistance against pest and diseases following PGPR is linked with the production of siderophore, HCN (hydrogen cyanide), antibiotics and increase in soil nutrient status and improvement in soil health (Vandana *et al.*, 2018). Nonetheless, a few concerns must be solved for these microbes for their long term uses. Moreover, AMF symbiosis also plays a significant role in plant performance; therefore, it should also keep in mind when using these microbes.

The screening and use of suitable pairing microbes in inadequate soils reduce the use of mineral fertilizers which improve the soil quality and production and quality of tea. Moreover, recognizing the AMF and other microbial induced stress tolerance and increase in crop production might help to further increase in crop production. Microbes and AMF must also be investigated at all levels to understand their functioning as bio-fertilizers and bio-control agents to improve agricultural productivity (Bag *et al.*, 2022). The rhizospheric engineering to promote positive plant microbe's interactions would also be a major concern in the future. However, rhizosphere engineering can create new opportunities to reduce the use of pesticides for sustaining environmental health. The recent biotechnological approaches such as can used to alter the expression of different genes. This would be a blueprint for microbes, commencing with the organism, finding genes, and producing the modified microbes that can be transferred into tea plants. This will surely improve the tea growth, yield and quality while it will reduce the reliance on chemical fertilizers.

Conclusions

Plant microbes have a significant potential to improve crop production and plant resistance against different stresses. Here in this review, we have discussed the role of various microbes in improve the growth, yield and quality of tea and the resistance of tea plants against different pests and diseases. Tea microbes improve nutrient cycling, water and nutrient uptake which in turn improves the yield of tea plants. Moreover, these microbes also improve the accumulation of nutrients amino acids, soluble proteins, flavonoids, catechuic acid, glucose, fructose, sucrose contents caffeine, polyphenols, bio-active compound concentration in tea plants. Moreover, these microbes also work as biofertilizers and fulfill nutrient needs which are a major advantage of these microbes. The use of pesticides and fertilizers is a fatal option it causes environmental and health issues. Therefore, microbes also offer the bio-control of various pest and diseases associated with tea which in turn improves the growth and yield of the tea plants. However, there are still many unanswered questions that should be addressed in future studies. Extensive research is needed to explore the mechanism associated with different microbes to improve the yield and quality of tea. Only limited microbes have been investigated

therefore it would be fascinating to conduct research on various microbes and their interaction with tea. The role of tea microbes must also explore in full detail against different pests and diseases attacking the tea plants. The genes governing microbial mediate growth and development in response to stress signaling must also be studied in future research. The recent improvement in sequencing and genome editing strategies identified the functions of various bacterial genes and these novel techniques can be used to alter the soil micro-biota which can enhance the tea growth, yield and resistance against pest and diseases. Besides this, the use of microbes can bring revolution in agriculture to fulfill the food needs of the raising population. Therefore, in the light of aforementioned facts research should be conducted to increase tea productivity while reducing the toxic effects of agro-chemicals for coming generations.

Authors' Contributions

Conceptualization; XZ; Writing - original draft: XZ; Writing - review and editing: HL, DH, MIA, JS, SL, AM, SL, JP, AP, MA and MUH.

All authors read and approved the final manuscript.

Ethical approval (for researches involving animals or humans)

Not applicable.

Acknowledgements

This research was funded by (2019086) and the Guangxi key R&D program (Guike AB19245039 and Guike AB22035057).

Conflict of Interests

The authors declare that there are no conflicts of interest related to this article.

References

- Abbasi H, Akhtar A, Sharf R (2015). Vesicular arbuscular mycorrhizal (VAM) fungi: a tool for sustainable agriculture. *American Journal of Plant Nutrition and Fertilization Technology* 5(2):40-49.
- Adesemoye AO, Kloepper JW (2009). Plant-microbes interactions in enhanced fertilizer-use efficiency. *Applied Microbiology and Biotechnology* 85(1):1-12. <https://doi.org/10.1007/s00253-009-2196-0>
- Ali M, Islam M, Saha N, Kanan AH (2014). Effects of microclimatic parameters on tea leaf production in different tea estates in Bangladesh. *World Journal of Agriculture and Soil Science* 10:134-140.
- Alurappa R, Chowdappa S, Narayanaswamy R, Sinniah UR, Mohanty, SK, Swamy MK (2018). Endophytic fungi and bioactive metabolites production: an update. *Microbial Biotechnology Springer, Singapore* 455-482. https://doi.org/10.1007/978-981-10-7140-9_21
- Anita S, Ponmurugan P, Ganesh BR (2012). Significance of secondary metabolites and enzymes secreted by *Trichoderma atroviride* isolates for the biological control of *Phomopsis canker* disease. *African Journal of Biotechnology* 11:10350-10357. <https://doi.org/10.5897/AJB12.599>

- Arafat Y, Ud Din I, Tayyab M, Jiang Y, Chen T, Cai Z, ... Lin S (2020). Soil sickness in aged tea plantation is associated with a shift in microbial communities as a result of plant polyphenol accumulation in the tea gardens. *Frontiers in Plant Science* 11:601. <https://doi.org/10.3389/fpls.2020.00601>
- Bag S, Mondal A, Banik A (2022). Exploring tea (*Camellia sinensis*) microbiome: Insights into the functional characteristics and their impact on tea growth promotion. *Microbiological Research* 254:126890. <https://doi.org/10.3389/fpls.2020.00601>
- Banik A, Chattopadhyay A, Ganguly S, Mukhopadhyay SK (2019a). Characterization of a tea pest specific *Bacillus thuringiensis* and identification of its toxin by MALDI-TOF mass spectrometry. *Industrial Crops and Products* 137:549-556. <https://doi.org/10.1016/j.indcrop.2019.05.051>
- Banik A, Dash GK, Swain P, Kumar U, Mukhopadhyay SK, Dangar TK (2019b). Application of rice (*Oryza sativa* L.) root endophytic diazotrophic *Azotobacter* sp. Strain Avi2 (MCC 3432) can increase rice yield under green house and field condition. *Microbiological Research* 219:56-65. <https://doi.org/10.1016/j.micres.2018.11.004>
- Banik A, Ganguly S, Mukhopadhyay, BB, Mukhopadhyay SK (2014). A new report on rapid, cheap and easily extractable mass spore production of *Beauveria bassiana* using recyclable polyurethane foams as support medium. *Journal of Microbiology and Biotechnology Research* 4:1-6. <https://doi.org/10.1016/j.micres.2021.126890>
- Barthakur BK (2011). Recent approach of Tocklai to plant protection in tea in North-east India. *Science and Culture* 77:381-384.
- Baslam M, Garmendia I, Goicoechea N (2013). The arbuscular mycorrhizal symbiosis can overcome reductions in yield and nutritional quality in greenhouse-lettuces cultivated at inappropriate growing seasons. *Scientia Horticulturae* 164:145-154. <https://doi.org/10.1016/j.scienta.2013.09.021>
- Begum N, Qin C, Ahanger MA, Raza S, Khan MI, Ashraf M, Zhang L (2019). Role of arbuscular mycorrhizal fungi in plant growth regulation: implications in abiotic stress tolerance. *Frontiers in Plant Science* 10:1068. <https://doi.org/10.3389/fpls.2019.01068>
- Benizri E, Baudoin E, Guckert A (2001). Root colonization by inoculated plant growth promoting rhizobacteria. *Biocontrol Science and Technology* 11(5):557-74. <https://doi.org/10.1080/09583150120076120>
- Bhardwaj D, Ansari MW, Sahoo RK, Tuteja N (2014). Biofertilizers function as key player in sustainable agriculture by improving soil fertility, plant tolerance and crop productivity. *Microbial Cell Factories* 13:66. <https://doi.org/10.1186/1475-2859-13-66>
- Bhattacharyya C, Banerjee S, Acharya U, Mitra A, Mallick I, Haldar A, ... Ghosh A (2020). Evaluation of plant growth promotion properties and induction of antioxidative defense mechanism by tea rhizobacteria of Darjeeling, India. *Scientific reports* 10(1):1-19. <https://doi.org/10.1038/s41598-020-72439-z>
- Bhattacharyya PN, Jha DK (2012). Plant growth-promoting rhizobacteria (PGPR): Emergence in agriculture. *World Journal of Microbiology and Biotechnology* 28:1327-1350. <https://doi.org/10.1007/s11274-011-0979-9>
- Bhattacharyya PN, Sarmah SR (2018). The role of microbes in tea cultivation. *Global Tea Science*. Burleigh Dodds Science Publishing, pp 155-188.
- Bhattacharyya PN, Sarmah SR, Dutta P, Tanti AJ (2015). Emergence in mapping microbial diversity in tea (*Camellia sinensis* (L.) O. Kuntze) soil of Assam, North-East India: A novel approach. *Emergence* 3(12).
- Bhattacharyya PN, Tanti B, Barman P, Jha DK (2014). Culture-independent metagenomic approach to characterize the surface and subsurface soil bacterial community in the Brahmaputra valley, Assam, North-East India, an Indo-Burma mega-biodiversity hotspot. *World Journal of Microbiology and Biotechnology* 30:519-28. <https://doi.org/10.1007/s11274-013-1467-1>
- Bora P, Bora LC (2021). Microbial antagonists and botanicals mediated disease management in tea, *Camellia sinensis* (L.) O. Kuntze: an overview. *Crop Protection* 105711. <https://doi.org/10.1016/j.cropro.2021.105711>
- Borah A, Das R, Mazumdar R, Thakur D (2019). Culturable endophytic bacteria of *Camellia* species endowed with plant growth promoting characteristics. *Journal of Applied Microbiology* 127(3):825-844. <https://doi.org/10.1111/jam.14356>
- Boruta T (2018). Uncovering the repertoire of fungal secondary metabolites: from Fleming's laboratory to the International Space Station. *Bioengineered* 9(1):12-16. <https://doi.org/10.1080/21655979.2017.1341022>
- Cakmakci R, Akçura S, Mustafa E (2021). Effect of co-inoculation of multi-traits bacteria based bio-formulations on the growth, yield and enzyme activities of tea. *Türk Tarım ve Doğa Bilimleri Dergisi* 8:594-604. <https://doi.org/10.30910/turkjans.807411>

- Çakmakçı R, Dönmez MF, Ertürk Y, Erat M, Haznedar A, Sekban R (2010). Diversity and metabolic potential of culturable bacteria from the rhizosphere of Turkish tea grown in acidic soils. *Plant and Soil* 332:299-318. <https://doi.org/10.1007/s11104-010-0295-4>
- Cernava T, Chen X, Krug L, Li H, Yang M, Berg G (2019). The tea leaf microbiome shows specific responses to chemical pesticides and biocontrol applications. *Science of Total Environment* 667:33-40. <https://doi.org/10.1016/j.scitotenv.2019.02.319>
- Chakraborty U, Chakraborty B, Basnet M (2006). Plant growth promotion and induction of resistance in *Camellia sinensis* by *Bacillus megaterium*. *Journal of Basic Microbiology* 46(3):186-195. <https://doi.org/10.1002/jobm.200510050>
- Chakraborty U, Chakraborty BN, Chakraborty AP (2012). Induction of plant growth promotion in *Camellia sinensis* by *Bacillus megaterium* and its bioformulations. *World Journal of Agricultural Sciences* 8(1):104-112.
- Chakraborty U, Chakraborty BN, Chakraborty AP, Sunar K, Dey PL (2013). Plant growth promoting rhizobacteria mediated improvement of health status of tea plants. *Indian Journal of Biotechnology* 12:20-31.
- Chandra R, Banik A (2021). Detoxification and bioconversion of arsenic and chromium. *Nanobiotechnology*. Elsevier, pp 253-270. <https://doi.org/10.1016/B978-0-12-822878-4.00016-X>
- Chelangat A, Gweyi-Onyango JP, Korir NK, Mwangi M (2021). Influence of arbuscular mycorrhizae on callusing and root colonization of tea (*Camellia sinensis*) clones in Kenya. *Asian Soil Research Journal* 5:21-26. <https://doi.org/10.9734/ASRJ/2021/v5i130098>
- Chen D, Ding Y, Ye H, Sun Y, Zeng X (2020). Effect of long-term consumption of tea (*Camellia sinensis* L.) flower polysaccharides on maintaining intestinal health in BALB/c mice. *Journal of Food Science* 85(6):1948-1955. <https://doi.org/10.1111/1750-3841.15155>
- Chen ZM, Lin Z (2015). Tea and human health: biomedical functions of tea active components and current issues. *Journal of Zhejiang University-Science B* 16:87-102. <https://doi.org/10.1631/jzus.B1500001>
- Chopra A, Vandana UK, Rahi P, Satpute S, Mazumder PB (2020). Plant growth promoting potential of *Brevibacterium sediminis* A6 isolated from the tea rhizosphere of Assam, India. *Biocatalysis and Agricultural Biotechnology* 27:101610. <https://doi.org/10.1016/j.bcab.2020.101610>
- Dash B, Soni R, Kumar V, Suyal DC, Dash D, Goel R (2019). Mycorrhizosphere: microbial interactions for sustainable agricultural production. *Mycorrhizosphere and Pedogenesis*. Springer, Singapore, pp 321-338. https://doi.org/10.1007/978-981-13-6480-8_18
- Dhar G, Mangar P, Saha A, Saha D (2018). Evaluation of the biocontrol efficacy of a *Serratia marcescens* strain indigenous to tea rhizosphere for the management of root rot disease in tea. *PLoS One* 13(2):e0191761. <https://doi.org/10.1371/journal.pone.0191761>
- Dodd IC, Ruiz-Lozano JM (2012). Microbial enhancement of crop resource use efficiency. *Current Opinion in Biotechnology* 23:236-242. <https://doi.org/10.1016/j.copbio.2011.09.005>
- Dutta BK, Kashyap MP, Morang P, Kumar D (2012). Growth promotion and bi-control approaches of brown root rot disease of tea by *Pseudomonas aeruginosa* (PM 105). *Journal of Plant Pathology and Microbiology*.
- Dutta J, Handique PJ, Thakur D (2015). Assessment of culturable tea rhizobacteria isolated from tea estates of Assam, India for growth promotion in commercial tea cultivars. *Frontiers in Microbiology* 6:1252. <https://doi.org/10.3389/fmicb.2015.01252>
- Erturk Y, Ercisli S, Sekban R, Haznedar A, Donmez MF (2008). The effect of plant growth promoting rhizobacteria (PGPR) on rooting and root growth of tea (*Camellia sinensis* var. *sinensis*) cuttings. *Romanian Biotechnological Letters* 13:3747-3756.
- Fang W, Yang L, Zhu X, Zeng L, Li X (2013). Seasonal and habitat dependent variations in culturable endophytes of *Camellia sinensis*. *Journal of Plant Pathology & Microbiology* 4(3):2157-471.
- Fauziah F, Seriwati MR, Pranoto E, Susilowati DN, Rachmiati Y (2019). Effect of indigenous microbes on growth and blister blight disease of tea plant. *Journal of Plant Protection Research* 529-534. <https://doi.org/10.24425/jppr.2019.131264>
- Fernando WGD, Nakkeeran S, Zhang Y (2005). Biosynthesis of antibiotics by PGPR and its relation in biocontrol of plant diseases. In: Siddiqui ZA (Ed). *PGPR: Biocontrol and Biofertilization*. Springer, Dordrecht, pp 67-109. https://doi.org/10.1007/1-4020-4152-7_3

- Gnanamangai BM, Ponnuragan P (2012). Evaluation of various fungicides and microbial based biocontrol agents against bird's eye spot disease of tea plants. *Crop Protection* 32:111-118. <https://doi.org/10.1016/j.cropro.2011.10.001>
- Greenlon A, Chang PL, Damtew ZM, Muleta A, Carrasquilla-Garcia N, Kim D, ... Patel JS (2019). Global-level population genomics reveals differential effects of geography and phylogeny on horizontal gene transfer in soil bacteria. *Proceedings of the National Academy of Science* 116(30):15200-15209. <https://doi.org/10.1073/pnas.1900056116>
- Guo S, Wang Q, Tang L, Zhang T, Li J, Xiao Y, Gao Y, Bai J, Xiao B, Gong C (2021). Inoculation with arbuscular mycorrhizal fungi reinforces tea plant's tolerance to salinity. *Journal of Plant Growth Regulation* 1-20. <https://doi.org/10.1007/s00344-021-10529-6>
- Gurusubramanian G, Borthakur M, Sarmah M, Rahman A (2005). Pesticide selection, precautions, regulatory measures and usage. In: Dutta AK, Gurusubramanian G, Barthakur BK (Eds). *Plant Protection in Tea*. Assam Printing Works Private Ltd., TTRI, TRA, Jorhat, Assam, India pp 81-91.
- Han L, Zhang H, Xu Y, Li Y, Zhou J (2021). Biological characteristics and salt-tolerant plant growth-promoting effects of an ACC deaminase-producing *Burkholderia pyrrocinia* strain isolated from the tea rhizosphere. *Archives of Microbiology* 1-12. <https://doi.org/10.1007/s00203-021-02204-x>
- Hazarika LK, Bhuyan M, Hazarika BN (2009). Insect pests of tea and their management. *Annual Review of Entomology* 54:267-284.
- Idris AL, Fan X, Muhammad MH, Guo Y, Guan X, Huang T (2020). Ecologically controlling insect and mite pests of tea plants with microbial pesticides: a review. *Archives of Microbiology* 1-10. <https://doi.org/10.1007/s00203-020-01862-7>
- Kakoki S, Kamimuro T, Tsuda K, Sakamaki Y (2018). Use of a lower-volume, surface pesticide spray conserves spider assemblages in a tea field. *Journal of Economic Entomology* 111(4):1595-1604. <https://doi.org/10.1093/jeet/toy147>
- Khan T, Mahbub MDA, Shawon M, Ali NM, Apu B, Tahmina I, Saha ML (2017). Rhizosphere associated bacteria and soil physicochemical properties of tea garden. *Bangladesh Journal of Botany* 46(4):1389-1396.
- Li Y, Li Z, Arafat Y, Lin W, Jiang Y, Weng B, Lin W (2017). Characterizing rhizosphere microbial communities in long-term monoculture tea orchards by fatty acid profiles and substrate utilization. *European Journal of Soil Biology* 81:48-54. <https://doi.org/10.1016/j.ejsobi.2017.06.008>
- Li YC, Li Z, Li ZW, Jiang YH, Weng BQ, Lin WX (2016). Variations of rhizosphere bacterial communities in tea (*Camellia sinensis* L.) continuous cropping soil by high-throughput pyrosequencing approach. *Journal of Applied Microbiology* 121(3):787-799. <https://doi.org/10.1111/jam.13225>
- Li Q, Chai S, Li Y, Huang J, Luo Y, Xiao L, Liu Z (2018). Biochemical components associated with microbial community shift during the pile-fermentation of primary dark tea. *Frontiers in Microbiology* 9:1509. <https://doi.org/10.3389/fmicb.2018.01509>
- Liu F, Weir BS, Damm U, Crous PW, Wang Y, Liu B, ... Cai L (2015). Unravelling *Colletotrichum* species associated with *Camellia*: employing ApMat and GS loci to resolve species in the *C. Gloeosporioides* complex. *Persoonia - Molecular Phylogeny and Evolution of Fungi* 35:63. <https://doi.org/10.3767/003158515X687597>
- Liu J, Xiao B, Wang L, Zhou X (2014). Influence of AMF on salt tolerance of tea. *Journal of Northwest A & F University-Natural Science Edition* 42(3):220-234.
- Liu J, Xiao B, Wang L, Li J, Pu G, Gao T, Liu W (2013). Influence of AM on the growth of tea plant and tea quality under salt stress. *Journal of Tea Science* 33:140-146.
- Mareeswaran J, Nepolean P, Jayanthi R, Premkumar SA, Radhakrishnan B (2015). In vitro studies on branch canker pathogen (*Macrophoma* sp.) infecting tea. *Journal of plant pathology and Microbiology* 6:284. <https://doi.org/10.4172/2157-7471.1000284>
- Mei L, Yang X, Zhang S, Zhang T, Guo J (2019). Arbuscular mycorrhizal fungi alleviate phosphorus limitation by reducing plant N: P ratios under warming and nitrogen addition in a temperate meadow ecosystem. *Science of Total Environment* 686:1129-1139. <https://doi.org/10.1016/j.scitotenv.2019.06.035>
- Mia M, Shamsuddin ZH, Wahab Z, Marziah M (2010). Effect of plant growth promoting rhizobacterial (PGPR) inoculation on growth and nitrogen incorporation of tissue-cultured *Musa* plantlets under nitrogen-free hydroponics condition. *AJCS* 4:85-90.
- Miransari M. (2011). Soil microbes and plant fertilization. *Applied Microbiology and Biotechnology* 92:875-885. <https://doi.org/10.1007/s00253-011-3521-y>

- Mishra DJ, Singh R, Mishra UK, Kumar SS (2013). Role of bio-fertilizer in organic agriculture: A review. *Research Journal of Recent Sciences* 2: 239-41.
- Mokhtar MM, El-Mougy NS (2014). Bio-compost application for controlling soil-borne plant pathogens - A review. *International Journal of Engineering Science and Technology* 4:61-68.
- Morang P, Dutta BK, Kumar, BS, Kashyap MP (2015). Growth promotion and bi-control approaches of brown root rot disease of tea by *Pseudomonas aeruginosa* (PM 105). *Plant Pathology & Microbiology* 3(5):1000129. <http://hdl.handle.net/123456789/2367>
- Mu D, Pan C, Qi Z, Qin H, Li Q, Liang K, ... Sun T (2021). Multivariate analysis of volatile profiles in tea plant infested by tea green leafhopper *Empoasca onukii* Matsuda. *Plant Growth Regulation* 95:111-120. <https://doi.org/10.1007/s10725-021-00729-1>
- Nath R, Sharma GD, Barooah M (2015). Plant growth promoting endophytic fungi isolated from tea (*Camellia sinensis*) shrubs of Assam, India. *Applied Ecology and Environmental Research* 13(3):877-891. https://doi.org/10.15666/aer/1303_877891
- Nepolean P, Jayanthi R, Pallavi RV, Balamurugan A, Kuberan T, Beulah T, Premkumar R (2012). Role of biofertilizers in increasing tea productivity. *Asian Pacific Journal of Tropical Biomedicine* 2(3):S1443-S1445. [https://doi.org/10.1016/S2221-1691\(12\)60434-1](https://doi.org/10.1016/S2221-1691(12)60434-1)
- Olanrewaju OS, Ayangbenro AS, Glick BR, Babalola OO (2019). Plant health: feedback effect of root exudates-rhizobiome interactions. *Applied Microbiology and Biotechnology* 103(3):1155-1166. <https://doi.org/10.1007/s00253-018-9556-6>
- Pallavi RV, Nepolean P, lamurugan A, Jayanthi R, Beulah T, Premkumar R (2012). *In vitro* studies of biocontrol agents and fungicides tolerance against grey blight disease in tea. *Asian Pacific Journal of Tropical Biomedicine* 2(1):S435-8. [https://doi.org/10.1016/S2221-1691\(12\)60202-0](https://doi.org/10.1016/S2221-1691(12)60202-0)
- Pandey A, Singh S, Palni LMS (2013). Microbial inoculants to support tea industry in India. *Indian Journal of Biotechnology* 12:13-19. <http://nopr.niscpr.res.in/handle/123456789/16484>
- Parada VD, Fuente DL, Landskron MK, Gonz'alez G, Quera MJ, Dijkstra R, ... Hermoso MA (2019). Short chain fatty acids (SCFAs)-mediated gut epithelial and immune regulation and its relevance for inflammatory bowel diseases. *Frontiers in Immunology* 10:277. <https://doi.org/10.3389/fimmu.2019.00277>
- Phour M, Sehrawat A, Sindhu SS, Glick BR (2020). Interkingdom signaling in plant-rhizomicrobiome interactions for sustainable agriculture. *Microbiological Research* 126589. <https://doi.org/10.1016/j.micres.2020.126589>
- Phukan I, Madhab M, Bordoloi M, Sarmah SR, Dutta P, Begum R, ... Barthakur BK (2012). Exploitation of PGP microbes of tea for improvement of plant growth and pest suppression: a novel approach. *Two and a Bud* 59(1):69-74.
- Premkumar R, Nepolean P, Vidhya Pallavi R, Balamurugan A, Jayanthi R (2012). Integrated disease management of grey blight in tea. *Two and a Bud* 59:27-30.
- Rajendran K, Devaraj P (2004). Biomass and nutrient distribution and their return of *Casuarina equisetifolia* inoculated with bio-fertilizers in farm land. *Biomass & Bioenergy* 26:235-249. <https://doi.org/10.1016/j.biombioe.2003.07.001>
- Rungsirivanich P, Supandee W, Futui W, Chumsai-Na-Ayudhya V, Yodsombat C, Thongwai N (2020). Culturable bacterial community on leaves of Assam tea (*Camellia sinensis* var. *assamica*) in Thailand and human probiotic potential of isolated *Bacillus* spp. *Microorganisms* 8(10):1585. <https://doi.org/10.3390/microorganisms8101585>
- Rustamova N, Bozorov K, Efferth T, Yili A (2020). Novel secondary metabolites from endophytic fungi: synthesis and biological properties. *Phytochemistry Reviews* 19(2):425-448. <https://doi.org/10.1007/s11101-020-09672-x>
- Saha D, Mukhopadhyay A (2013). Insecticide resistance mechanisms in three sucking insect pests of tea in reference to North-East India; An appraisal. *International Journal of Tropical Insect Science* 33(1):46-70. <https://doi.org/10.1017/S1742758412000380>
- Sanmartín C, Garmendia I, Romano B, Díaz M, Palop JA, Goicoechea N (2014). Mycorrhizal inoculation affected growth, mineral composition, proteins and sugars in lettuces biofortified with organic or inorganic selenocompounds. *Scientia Horticulturae* 180:40-51. <https://doi.org/10.1016/j.scienta.2014.09.049>
- Saravanakumar D, Vijayakumar C, Kumar N, Samiyappan R (2007). PGPR induced defense responses in the tea plant against blister blight disease. *Crop Protection* 26:556-565. <https://doi.org/10.1016/j.cropro.2006.05.007>

- Shan W, Zhou Y, Liu H, Yu X (2018). Endophytic actinomycetes from tea plants (*Camellia sinensis*): isolation, abundance, antimicrobial, and plant-growth-promoting activities. *BioMed Research International* 2018. <https://doi.org/10.1155/2018/1470305>
- Shang J, Liu B (2021). Application of a microbial consortium improves the growth of *Camellia sinensis* and influences the indigenous rhizosphere bacterial communities. *Journal of Applied Microbiology* 130(6):2029-2040. <https://doi.org/10.1111/jam.14927>
- Shao YD, Hu XC, Wu QS, Yang TY, Srivastava AK, Zhang DJ, Kuca K (2021). Mycorrhizas promote P acquisition of tea plants through changes in root morphology and P transporter gene expression. *South African Journal of Botany* 137:455-462. <https://doi.org/10.1016/j.sajb.2020.11.028>
- Shao YD, Zhang DJ, Hu XC, Wu QS, Jiang CJ, Gao XB, Kuca K (2019). Arbuscular mycorrhiza improves leaf food quality of tea plants. *Notulae Botanicae Horti Agrobotanici Cluj-Napoca* 47(3).
- Shao YD, Zhang DJ, Hu XC, Wu QS, Jiang CJ, Xia TJ, ... Kuča K (2018). Mycorrhiza-induced changes in root growth and nutrient absorption of tea plants. *Plant, Soil and Environment* 64(6):283-289. <https://doi.org/10.17221/126/2018-PSE>
- Sharma D, Kayang H (2017). Effects of arbuscular mycorrhizal fungi (AMF) on *Camellia sinensis* (L.) O. Kuntze under greenhouse conditions. *Journal of Experimental Biology* 5:235-241. [http://dx.doi.org/10.18006/2017.5\(2\).235.241](http://dx.doi.org/10.18006/2017.5(2).235.241)
- Sharma E, Joshi R, Gulati A (2018). l-Theanine: an astounding sui generis integrant in tea. *Food Chemistry* 242:601-610. <https://doi.org/10.1016/j.foodchem.2017.09.046>
- Singh S, Pandey A, Kumar B, Palni LMS (2010). Enhancement in growth and quality parameters of tea [*Camellia sinensis* (L.) O. Kuntze] through inoculation with arbuscular mycorrhizal fungi in an acid soil. *Biology and Fertility of Soils* 46:427-433. <https://doi.org/10.1007/s00374-010-0448-x>
- Sowndhararajan K, Marimuthu S, Manian S (2013). Integrated control of blister blight disease in tea using the biocontrol agent, *Ochrobactrum anthropi* strain BMO-111 with chemical fungicides. *Journal of Applied Microbiology* 114:1491-1499. <https://doi.org/10.1111/jam.12159>
- Trabelsi D, Mhamdi R (2013). Microbial inoculants and their impact on soil microbial communities: a review. *BioMed Research International*. <https://doi.org/10.1155/2013/863240>
- Tshikhudo PP, Ntushelo K, Mudau FN, Salehi B, Sharifi-Rad M, Martins N, Sharifi-Rad J (2019). Understanding *Camellia sinensis* using omics technologies along with endophytic bacteria and environmental roles on metabolism: a review. *Applied Sciences (Basel)* 9(2):281. <https://doi.org/10.3390/app9020281>
- Vandana UK, Chopra A, Choudhury A, Adapa D, Mazumder PB (2018). Genetic diversity and antagonistic activity of plant growth promoting bacteria, isolated from tea-rhizosphere: a culture dependent study. *Biomedical Research* 29(4):853-864.
- Vessey JK (2003). Plant growth promoting rhizobacteria as biofertilizers. *Plant and Soil* 255:571-86. <https://doi.org/10.1023/A:1026037216893>
- Wan X (2003). *Tea Biochemistry*, 3rd Edn, Vol. 76. Beijing: China Agriculture Press, pp 185.
- Wang B (2019). Study on the correlation between microorganism and quality formation of pu'er tea during fermentation. In: *IOP Conference Series: Earth and Environmental Science*: IOP Publishing), 032055. <https://iopscience.iop.org/journal/1755-1315>
- Wang M, Deng B, Fu X, Sun H, Xu Z (2019). Characterizations of microbial diversity and machine oil degrading microbes in machine oil contaminated soil. *Environmental Pollution* 255:113190. <https://doi.org/10.1016/j.envpol.2019.113190>
- Wang M, Jiang P (2015). Colonization and diversity of AM fungi by morphological analysis on medicinal plants in southeast China. *Transfusion and Apheresis Science* 2015. <https://doi.org/10.1155/2015/753842>
- Wang M, Sun H, Xu L, Xu Z (2021). Bacterial diversity in tea plant (*Camellia sinensis*) rhizosphere soil from Qinling Mountains and its relationship with environmental elements. *Plant and Soil* 460(1):403-415. <https://doi.org/10.1007/s11104-020-04822-8>
- Wardle DA, Bardgett RD, Klironomos JN, Setälä H, van der Putten WH, Wall DH (2004). Ecological linkages between above ground and below ground biota. *Science* 304:1629-33. <https://doi.org/10.1126/science.1094875>
- Win PM, Matsumura E, Fukuda K (2017). Diversity of tea endophytic fungi: cultivar-and tissue preferences. *Applied Ecology and Environmental Research* 16:677-695. http://dx.doi.org/10.15666/aeer/1601_677695

- Wu QS, Shao YD, Gao XB, Xia TJ, Kuřca K (2019). Characterization of AMF-diversity of endosphere versus rhizosphere of tea (*Camellia sinensis*) crops. *Indian Journal of Agricultural Sciences* 89(2):348-352.
- Xie H, Feng X, Wang M, Wang Y, Kumar Awasthi, M, Xu P (2020). Implications of endophytic microbiota in *Camellia sinensis*: a review on current understanding and future insights. *Bioengineered* 11(1):1001-1015. <https://doi.org/10.1080/21655979.2020.1816788>
- Yan X, Wang Z, Mei Y, Wang L, Wang X, ... Wei C (2018). Isolation, diversity, and growth-promoting activities of endophytic bacteria from tea cultivars of Zijuan and Yunkang-10. *Frontiers in Microbiology* 9:1848. <https://doi.org/10.3389/fmicb.2018.01848>
- Yu Y, Zhang J, Huang C, Hou X, Sun X, Xiao B (2018). Reference genes selection for quantitative gene expression studies in tea green leafhoppers, *Empoasca onukii* Matsuda. *PLoS One* 13(10):e0205182. <https://doi.org/10.1371/journal.pone.0205182>
- Zhao QH, Sun LT, Wang Y, Ding ZT, Li M (2014). Effects of arbuscular mycorrhizal fungi and nitrogen regimes on plant growth, nutrient uptake, and tea quality in *Camellia sinensis* (L.) O. Kuntze. *Journal of Plant Physiology* 50(2):164-170.
- Zubek S, Rola K, Szweczyk A, Majewska ML, Turnau K (2015). Enhanced concentrations of elements and secondary metabolites in *Viola tricolor* L. Induced by arbuscular mycorrhizal fungi. *Plant Soil* 390(1):129-142. <https://doi.org/10.1007/s11104-015-2388-6>



The journal offers free, immediate, and unrestricted access to peer-reviewed research and scholarly work. Users are allowed to read, download, copy, distribute, print, search, or link to the full texts of the articles, or use them for any other lawful purpose, without asking prior permission from the publisher or the author.



License - Articles published in *Notulae Botanicae Horti Agrobotanici Cluj-Napoca* are Open-Access, distributed under the terms and conditions of the Creative Commons Attribution (CC BY 4.0) License. © Articles by the authors; Licensee UASVM and SHST, Cluj-Napoca, Romania. The journal allows the author(s) to hold the copyright/to retain publishing rights without restriction.

Notes:

- **Material disclaimer:** The authors are fully responsible for their work and they hold sole responsibility for the articles published in the journal.
- **Maps and affiliations:** The publisher stay neutral with regard to jurisdictional claims in published maps and institutional affiliations.
- **Responsibilities:** The editors, editorial board and publisher do not assume any responsibility for the article's contents and for the authors' views expressed in their contributions. The statements and opinions published represent the views of the authors or persons to whom they are credited. Publication of research information does not constitute a recommendation or endorsement of products involved.