

Modelling plant morphometric parameters as predictors for successful cultivation of some medicinal *Agastache* species

Rodica VÂRBAN¹, Roxana VIDICAN¹, Andreea D. ONA¹,
Dan VÂRBAN¹, Andrei STOIE^{1,2*}, Ștefania GÂDEA¹, Sorin VÂTCĂ¹,
Valentina STOIAN¹, Ioana CRIȘAN^{1,2}, Vlad STOIAN¹

¹University of Agricultural Sciences and Veterinary Medicine Cluj-Napoca, Faculty of Agriculture, Calea Mănăștur 3-5, 400372 Cluj-Napoca, Romania; rodica.varban@usamvcluj.ro; roxana.vidican@usamvcluj.ro; andreea.ona@usamvcluj.ro; dan.varban@usamvcluj.ro; stefania.gadea@usamvcluj.ro; sorin.vatca@usamvcluj.ro; valentina.stoian@usamvcluj.ro; ioana.crisan@usamvcluj.ro; vlad.stoian@usamvcluj.ro

²University of Agricultural Sciences and Veterinary Medicine Cluj-Napoca, Agro-Botanical Garden, Calea Mănăștur 3-5, 400372 Cluj-Napoca, Romania; andrei.stoie@usamvcluj.ro (*corresponding author)

Abstract

Researches carrying evidence for various uses and bioactive principles of *Agastache* spp. are justifying the upscaling into cultivation of these medicinal species. But, hindrances in their cultivation exist due to the insufficient documentation of their biology under field conditions. Because productivity of these medicinal species (*herba*) is ensured by the combined contribution of plant agronomic traits, these are related to the feasibility of the crop and therefore, can be used as predictors for successful cultivation. The aim of this study was to evaluate comparatively four valuable *Agastache* species (*A. mexicana*, *A. scrophulariifolia*, *A. foeniculum*) and one cultivar (*A. rugosa* 'After Eight'), in order to identify the favourability for cultivation in local conditions (Romania). Based on the structural indicators of plant morphology (plant height, shoot number, leaf number, leaf length and width, inflorescence length, verticillaster number and flower number), registered over the span of two years, were explored relationships and similarities as well as their implications in previsioning the phenotypic potential. The results showed that studied species acclimatized successfully and all agronomic parameters studied increased in values in the second year. The average plant height in second year (2020) was 109.8 cm and average inflorescences length 9.6 cm. Stable positive correlations between inflorescence length with plant height and shoot number were observed, while differences among species became pronounced as plants become established, evidenced by clearer distinction in the second year. Phenotypic potential in the absence of inputs enables the feasibility assessment for medicinal plants introduced for cultivation in new regions.

Keywords: agronomic trait; correlation; field conditions; *herba Agastache*

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Introduction

The importance of medicinal plants today extends beyond the traditional uses because these represent trade commodities that cover the demand of diverse industry and market sectors (Schippmann *et al.*, 2002). Worldwide the trade in medicinal plants, herbal drugs and raw materials experiences a growth rate of 15% per year, while about 25% of modern medicines are derived from plants (Ahmad and Ahmad, 2019). At global level it is estimated that production of medicinal and aromatic plants exceeds 300 million tons, but it is difficult to assess how many are actually traded. In Europe there are over 30 thousand companies dealing in this sector, and an area exceeding 200 thousand hectares cultivated Europe-wide, with countries such as France, Poland, Spain, Bulgaria, Croatia and Czech Republic as top players. However, current area cultivated with medicinal plants covers only a small fraction of the industry needs (Argyropoulos, 2019). The number of plant species used for medicinal purposes world-wide might exceed 50,000, but in Europe about 2,000 plant species are in trade for medicinal use (Schippmann *et al.*, 2006).

There is an increasing demand for medicinal plants for the production of plant-based medicines and drugs, various plant-based health products, food supplements, cosmetics and plant protection products (Li and Weng, 2017; Ahmad and Ahmad, 2019; Argyropoulos, 2019). Major markets are located in developed countries, with Europe as main import region due to the large demand for herbal raw materials that end up in various processing or food sectors (Schippmann *et al.*, 2002; Argyropoulos, 2019). Medicinal plants used worldwide are collected from the wild, grown in home gardens or cultivated either as sole cropping or in intercropping systems. There is preference for cultivated material, because most herbal companies, mass-market and pharmaceutical companies opt for cultivated raw material (Schippmann *et al.*, 2002). Also, research has shown that among committed consumers there is a preference for quality, organically-grown plants, ecological certification and traceability (Cadar *et al.*, 2021). Growers can consider extending their range of cultivated species to take advantage of the industry and market opportunities. This can be achieved by extending the cultivation of medicinal species and introducing novel ones into cultivation (Schippmann *et al.*, 2002, 2006; Argyropoulos, 2019).

Agastache is a genus from the family Lamiaceae that includes aromatic species native to North America and East Asia. Plants have prevalently ovate or deltoid-ovate leaf shapes with serrate or crenate margins. The inflorescence is comprised by small flowers arranged in verticillasters (Lint and Epling, 1945; WFO, 2021). They exhibit a hemicryptophyte life-form in conditions of Romania (Vârban *et al.*, 2021). The genus does not occur naturally in Europe, but species from this genus are promising medicinal plants from which *herba* is harvested during flowering and can be valorised as green or dried biomass as well as for essential oil production (Duda *et al.*, 2013; Muntean *et al.*, 2016). Numerous studies regarding the importance of these medicinal species justifies their cultivation: for ornamental purposes (Lord, 2003; Marchioni *et al.*, 2021), as melliferous plant (Anand *et al.*, 2018; Anand *et al.*, 2019a; Anand *et al.*, 2019b), for foods and drinks (Fuentes-Granados *et al.*, 1998; Carović-Stanko *et al.*, 2016; Quattrocchi, 2016; Najjar *et al.*, 2019; Marchioni *et al.*, 2021), for valuable essential oil (Ivanonv *et al.*, 2019; Gonzalez-Ramírez *et al.*, 2021) and for source of specific bioactive compounds (Zielińska and Matkowski, 2014; Quattrocchi, 2016; Hwang *et al.*, 2021). But while biochemical properties have received attention (Kovalenko *et al.*, 2019; Najjar *et al.*, 2019; Yeo *et al.*, 2021; Najafi *et al.*, 2022) there is a lack of information that could consolidate technological packages to enable the successful cultivation of species from this genus (Palma-Tenango *et al.*, 2021).

Agencies advise for moderation in regards with inputs for medicinal crops (EMEA, 2006). Particularly, the potential accumulation of nitrites and nitrates in medicinal plants is one of the most concerning (Mohamed, 2012; Özcan and Akbulut, 2008; Nchu *et al.*, 2017), and can occur due to excessive N fertilization (Nchu *et al.*, 2017). Therefore, it is preferable that productivity of medicinal plants is not highly reliant on application of fertilizers or stimulators to enhance yield. But in these conditions, feasibility of the crop becomes dependent of the natural phenotypic expression and as a consequence the precise knowledge about the phenotypic potential of the plants has high agronomic relevance.

An outlook on morphometric specifications for *Agastache* species from the existing body of literature (Table 1), reveals that in many studies the measurements were performed on plants grown under controlled conditions and in most cases the plants were not observed until advanced stages of growth (Khorsandi *et al.*, 2010; Do *et al.*, 2020; Jahani *et al.*, 2018; Kim *et al.*, 2018; Lam, *et al.*, 2020a; Lam *et al.*, 2020b). At the same time, some studies focused only on some reproductive traits of plants (Jang *et al.*, 2015; Rudik, 2016). The studies that cover the whole range of morphometric parameters representing key agronomic traits linked to productivity of these species, are few (Ok and Chae, 1998; Lee *et al.*, 1999; Sheahan, 2012; Melnychuk and Rakhmetov, 2016; Carrillo-Galván *et al.*, 2020). As a consequence, the information obtained is insufficient for the optimization of field cultivation of *Agastache* species.

Table 1. Review of morphometric specifications for *Agastache* species studied

Spp.	Condi-tions ¹	Details of the context	Morphometric ²		Sources
			Veg.	Repr.	
<i>A. foeniculum</i>	G	Comparative screening under hydroponic culture	x	-	(Do <i>et al.</i> , 2020)
	G	Effect of ploidy level on morphometric traits	x	x	(Talebi <i>et al.</i> , 2017)
	G	Influence of foliar stimulants/fertilizers on plant growth	x	-	(Jahani <i>et al.</i> , 2018)
	F	Acclimatization in Kremenets Botanical Garden	x	x	(Melnychuk and Rakhmetov, 2016)
	I	Direct <i>in vitro</i> plant regeneration	x	-	(Hosseini and Moharrami, 2014)
	G	Salinity and plant growth	x	-	(Khorsandi <i>et al.</i> , 2010)
	F	Description - Gardening encyclopedia	x	-	(Lord, 2003)
	F	Description - Botany and taxonomic keys	x	x	(Lint and Epling, 1945)
<i>A. mexicana</i>	F	Genotypes at different stages of domestication	x	x	(Carrillo-Galván <i>et al.</i> , 2020)
	I	<i>In vitro</i> regeneration	x	-	(Carmona-Castro <i>et al.</i> , 2019)
	G	Effect of ploidy level on morphometric traits	x	x	(Talebi <i>et al.</i> , 2017)
	G	Influence of foliar stimulants/fertilizers on plant growth	x	-	(Jahani <i>et al.</i> , 2018)
	F	Acclimatization in Kremenets Botanical Garden	x	x	(Melnychuk and Rakhmetov, 2016)
	F	Description - Gardening encyclopedia	x	x	(Lord, 2003)
	F	Description - Botanic and taxonomic keys	x	x	(Lint and Epling, 1945)
<i>A. rugosa</i>	G	Influence of root zone temperature on plant growth in hydroponic culture	x	-	(Lam <i>et al.</i> , 2020a)
	G	Influence of electrical conductivity and plant growth in hydroponic culture	x	-	(Lam <i>et al.</i> , 2020b)
	G	Screening varieties in hydroponic culture	x	-	(Do <i>et al.</i> , 2020)
	G	Influence of electrical conductivity on plant growth in hydroponic culture	x	-	(Kim <i>et al.</i> , 2018)
	F	Comparative inflorescence morphology	-	x	(Rudik, 2016)
	F	Floral dimorphism in South Korean natural populations	-	x	(Jang <i>et al.</i> , 2015)
	I	<i>In vitro</i> propagation of shoots	x	-	(Zielińska <i>et al.</i> , 2011)
	F	Influence of N fertilization on plant growth	x	x	(Ohk <i>et al.</i> , 2000)
	F	Description - Gardening encyclopedia	x	-	(Lord, 2003)

	F	Comparison of some genotypes from different regions of South Korea	x	x	(Lee <i>et al.</i> , 1999)
	F	Comparison of some South Korean accessions	x	x	(Ok and Chae, 1998)
	N/A	Influence of seeding dates, density and fertilizer on agronomic traits	x	-	(Choi and Seo, 1993)
	F	Description - Botany and taxonomic keys	x	x	(Lint and Epling, 1945)
<i>A. scrophulariifolia</i>	I	Germination and regeneration of plants from nodal segments on different media	x	-	(Polivanova and Cherednichenko, 2017)
	F	Plant fact sheet (United States Department of Agriculture)	x	x	(Sheahan, 2012)
	F	Conservation and Research Plan for New England, United States of America	x	x	(Corrigan, 2002)
	F	Description - Botany and taxonomic keys	x	x	(Lint and Epling, 1945)

*Notes (legend): ¹Conditions of cultivation: G - controlled environment (greenhouse, hydroponic, pots); F - field (outdoors garden, crop field or natural populations); I - *in vitro*; N/A - undetermined (paper in other language and not specified in the English abstract). ²Morphometric specifications: Vegetative (veg.) - one or more characteristics, such as plant height and spread, shoot, root and leaf; Reproductive (rep.) - one or more characteristics of inflorescence and/or flowers (including flower parts).

In previous work we demonstrated that a first step for defining suitability for cultivation is to determine the phenology in local conditions (Vârban *et al.*, 2021). This is essential particularly for perennial species from genus *Agastache*, since depending of their place of origin might or might not be hardy in local climate. Results have shown that *Agastache* species studied resisted over winter in Cluj-Napoca, Romania. Moreover, phenology revealed that the succession of phenophases follows similar trend, allowing their cultivation at least in Transylvania region (Vârban *et al.*, 2021). The current study concerned with answering the next question that growers might have: what is the phenotypic potential of these species? The answer of this question might allow growers to choose those species that they consider best for their needs.

The aim of the research was to evaluate comparatively the phenotypic potential of some valuable *Agastache* species by assessing plant morphometric characteristics under field conditions. All the characters studied are of agronomic interest, determining the production and quality of the biological product obtained. For defining the morphometric parameters with highest precision, in the experimental field were not performed interventions (irrigation, fertilization, growth stimulators etc.). The research attempts to offer proof of concept for facile analysis of agronomic traits for predicting favourability of cultivation of some promising medicinal species. The results could complete a gap from literature by providing specifications regarding phenotypic expression of four *Agastache* species (*A. foeniculum*, *A. rugosa*, *A. mexicana*, *A. scrophulariifolia*) and a cultivar (*A. rugosa* 'After Eight') under field conditions. Two objectives were defined: evaluation of morphometric parameters with agronomic importance and study of the relationships between them; assessment of similarity between species based on projection of the morphometric traits.

Materials and Methods

Description of the study site

The experiment took place in the years 2019 and 2020 in the Agro-Botanical Garden of the University of Agricultural Sciences and Veterinary Medicine (UASVM) from Cluj-Napoca, Romania and located at latitude 46°45'36" N and longitude 23°34'24" E, elevation 380-430 m (Index Seminum - Hortus Agro-Botanicus, 2021).

The local climate is temperate-continental with oceanic influence (Criveanu, 2001) and Dfb (warm humid continental) according to Köppen-Geiger classification (<https://en.climate-data.org/>). According to

climatic conditions from the experimental interval registered by the UASVM Cluj-Napoca weather station, in the year 2020 precipitations were more abundant, and were associated with lower average temperatures over summer months, compared with year 2019 (Figure 1).

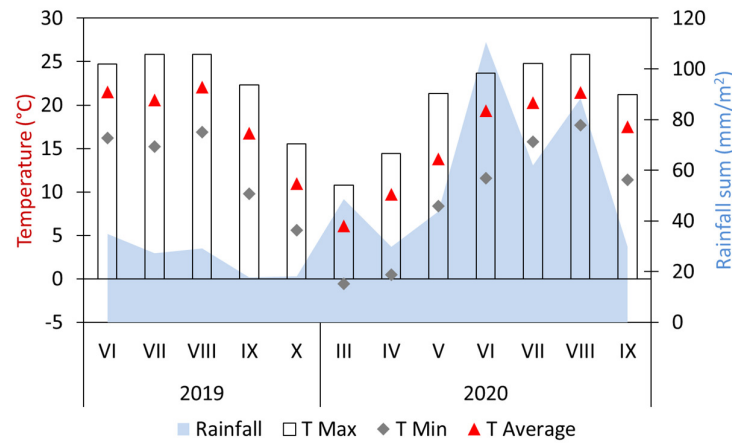


Figure 1. Climatic conditions during plant development in the field, according to weather station of UASVM Cluj-Napoca, Romania

In the Agro-Botanical Garden, the soil has a loam-clay texture, pH 6.72, and low humus content (1.35%). The supply with macronutrients is good: Nitrogen 0.461%, Potassium 312 ppm, and Phosphorus 68 ppm (Vârban *et al.*, 2021).

Biological material

In the experiment were studied accessions of four species of *Agastache* and one cultivar obtained from Botanisches Institut und Botanischer Garten from Universität Gesamthochschule Essen, Germany (Table 2). Voucher specimens are stored at the Agro-Botanical Garden Herbarium from UASVM Cluj-Napoca, 30078–30082 voucher numbers. These species have not been cultivated before in local conditions, with the exception of *A. foeniculum*.

Table 2. The *Agastache* sp. accessions from the study

Species/Cultivar	International Plant Exchange Number	Native range (Vârban <i>et al.</i> , 2021)
<i>A. foeniculum</i> (Pursh) Kuntze	XX-0-CLA-4152	throughout North America
<i>A. scrophulariifolia</i> (Willd.) Kuntze	XX-0-CLA-4155	throughout North America
<i>A. mexicana</i> (Kunth) Lint et Epling	XX-0-CLA-4153	southern part of North America (Mexico)
<i>A. rugosa</i> (Fisch. et C.A.Mey.) Kuntze	XX-0-CLA-4154	throughout East Asia
<i>A. rugosa</i> ‘After Eight’	XX-0-CLA-1950	-

Experimental procedures

The cultivation has been described in our previous work (Vârban *et al.*, 2021) and therefore only briefly reminded here. The field crop was started in June 2019 by planting seedlings obtained in greenhouse during the spring. These species are perennials, and once planted the species were monitored for two consecutive years. There were no plant losses during winter, all plants resisted over winter. After establishment of the crop, in this experiment there were no interventions such as irrigation, fertilization, growth stimulators or application of plant protection products.

The experimental plot had 1000 m² and organized in a subdivided plots design.

Sampling design

The parameters were quantified at phenophase stage 7 on BBCH scale, in both of the experimental years (Vârban *et al.*, 2021). This coincides with the moment when plants were flowering and the growth of the shoot has stopped. During this phenophase, plants are also usually harvested for *herba*. The observations were conducted concomitantly with phenology (Vârban *et al.*, 2021), during the same experimental years: 2019 and 2020. Determination of morphometric traits completes the assessment for favourability of cultivation, as a second step after phenological assessment (Vârban *et al.*, 2021).

Determinations were conducted for ten plants of each species in four replicates each year, randomly selected (in total 200 plants were analysed each year).

The agronomic traits determined were:

- Vegetative traits: plant height (cm) (PH), shoot number per plant (SN), leaf length (cm) (LL), leaf width (cm) (LW), leaf number per shoot (LN);
- Reproductive traits: inflorescence length (cm) (IL), verticillaster number per inflorescence (VN), flower number per verticillaster (FN).

A number of ten determinations for each agronomic trait were obtained from every plant analysed. The leaf number per shoot was determined by counting all normally and fully developed leaves. To determine the size of the leaves, leaves located at different levels on the plant (base, middle and upper third of the shoots) were harvested and measured with the ruler. The inflorescence length was determined by measuring the inflorescence with the ruler. The ten inflorescences from each plant analysed were taken from different levels on the plant. The number of flowers per verticillaster was determined by counting the flowers in each verticillaster of the inflorescence.

The studied species and method of determination for some parameters are illustrated in Figure 2.

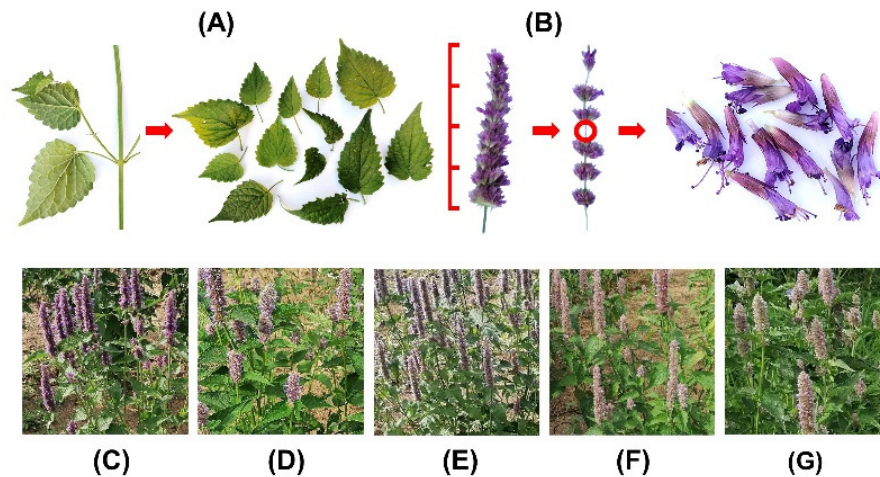


Figure 2. Method of quantifying some quantitative traits (A-B) in some *Agastache* species/cultivar: (C) *A. scrophulariifolia*; (D) *A. rugosa*; (E) *A. foeniculum*; (F) *A. rugosa* 'After Eight'; (G) *A. mexicana*

Statistical procedures

The tests applied were chosen in accordance with the objectives of the study.

For the comparative assessment of agronomic traits, were applied ANOVA test followed by Tukey's (HSD) honestly significant differences test using RStudio version 1.4.1106 (<https://www.rstudio.com>) for R statistical software (<https://www.r-project.org>). For these tests were used the packages "agricolae" (Mendiburu, 2020) and "broom" (Robinson *et al.*, 2021).

Relationships between agronomic traits were studied by using Pearson correlations. The correlations coefficients were obtained using package "Hmisc" (Harrel and Dupont, 2021) in R (<https://www.r->

project.org). Visual representation of correlograms was obtained with PAST software version 4.05 (Hammer, 2021).

Previsioning of the interrelationships between parameters was achieved by comparing models fitted by maximum likelihood with Akaike's Information Criterion (AIC) within the "vegan" package for R (Oksanen *et al.*, 2020), and verified with "mass" package (Ripley *et al.*, 2021). The resulted regressions were verified and completed with formulas from package "caret" (Kuhn *et al.*, 2021).

Projection of similarity between the studied species based on morphometric parameters was achieved through Principal Component Analysis (PCA) using PAST software version 4.05 (Hammer, 2021).

Results

Comparative evaluation of agronomic traits

Analysis of variance revealed that all agronomic traits were highly influenced by species/cultivar ($p < 0.001$) (Table 3).

Between the two experimental years, all of the eight agronomic traits studied increased in value. All species exceeded one meter in height in the second year. The cultivar *A. rugosa* 'After Eight' had in both years the tallest plants with longest inflorescence.

Table 3. Agronomic traits of *Agastache* sp. studied in Cluj-Napoca in the experimental years (mean \pm SE)

Year	Species	Plant height (cm)	Shoot number	Leaf length (cm)	Leaf width (cm)	Leaf number	Inflorescence length (cm)	Verticillaster number	Flower number
2019	<i>A. rugosa</i> 'After Eight'	85.0 \pm 0.85 a	6.7 \pm 0.13 a	6.0 \pm 0.22 b	4.0 \pm 0.21 b	24.0 \pm 1.04 b	11.2 \pm 0.38 a	10.6 \pm 0.36 a	24.0 \pm 0.96 b
	<i>A. foeniculum</i>	70.0 \pm 1.08 c	4.5 \pm 0.22 c	4.5 \pm 0.15 c	3.4 \pm 0.16 c	23.1 \pm 0.88 b	7.8 \pm 0.25 b c	8.5 \pm 0.36 b	26.8 \pm 1.19 b
	<i>A. mexicana</i>	65.0 \pm 1.03 d	5.6 \pm 0.29 b	5.8 \pm 0.25 b	3.5 \pm 0.18 bc	27.3 \pm 1.00 a	7.0 \pm 0.29 c	12.0 \pm 0.58 a	36.2 \pm 1.06 a
	<i>A. rugosa</i>	65.3 \pm 0.63 d	6.3 \pm 0.16 ab	5.8 \pm 0.04 b	4.6 \pm 0.02 a	19.8 \pm 0.34 c	8.0 \pm 0.07 b	9.0 \pm 0.24 b	24.6 \pm 0.48 b
	<i>A. scrophulariifolia</i>	73.6 \pm 0.74 b	4.0 \pm 0.11 c	7.5 \pm 0.05 a	5.0 \pm 0.06 a	25.0 \pm 0.70 ab	7.5 \pm 0.09 b c	8.5 \pm 0.12 b	17.8 \pm 0.30 c
	<i>F</i> test	58.86	34.71	41.24	22.37	10.95	45.05	17.73	58.86
	<i>p</i> value	$p < 0.001$	$p < 0.001$	$p < 0.001$	$p < 0.001$	$p < 0.001$	$p < 0.001$	$p < 0.001$	$p < 0.001$
2020	<i>A. rugosa</i> 'After Eight'	119.6 \pm 0.60 a	12.4 \pm 0.19 b	7.0 \pm 0.18 b	4.8 \pm 0.11 b	46.4 \pm 0.39 b	13.2 \pm 0.27 a	13.2 \pm 0.19 b	34.6 \pm 0.64 b
	<i>A. foeniculum</i>	104.0 \pm 1.74 c	9.4 \pm 0.30 c	5.2 \pm 0.16 c	4.0 \pm 0.12 c	44.3 \pm 0.93 b	8.8 \pm 0.19 b c	12.0 \pm 0.39 b	36.6 \pm 0.58 b
	<i>A. mexicana</i>	103.5 \pm 0.85 c	9.7 \pm 0.25 c	6.8 \pm 0.15 b	4.8 \pm 0.12 b	54.1 \pm 0.75 a	8.3 \pm 0.20 c	15.2 \pm 0.34 a	42.2 \pm 0.75 a
	<i>A. rugosa</i>	107.0 \pm 0.35 c	13.7 \pm 0.34 a	6.8 \pm 0.04 b	5.6 \pm 0.04 a	37.6 \pm 0.38 d	8.7 \pm 0.05 b c	13.3 \pm 0.26 b	35.4 \pm 0.53 b
	<i>A. scrophulariifolia</i>	115.0 \pm 1.12 b	8.3 \pm 0.22 d	7.6 \pm 0.05 a	5.7 \pm 0.04 a	40.0 \pm 0.40 c	9.3 \pm 0.16 b	10.6 \pm 0.20 c	22.8 \pm 0.41 c
	<i>F</i> test	46.67	72.00	44.51	52.52	109.38	118.81	35.96	143.04
	<i>p</i> value	$p < 0.001$	$p < 0.001$	$p < 0.001$	$p < 0.001$	$p < 0.001$	$p < 0.001$	$p < 0.001$	$p < 0.001$

*Notes (legend): Different letters denote significant differences (Tukey HSD significance test)

By comparison, *A. rugosa* maintained in both years lowest leaf number per shoot. *A. scrophulariifolia* presented in both years the widest and longest leaves. *A. mexicana* presented in both years the shortest plants, but highest leaf number, highest number of verticillasters per inflorescence and highest flower number per verticillaster in both years. *A. foeniculum* did not present any top performance in either of the two years compared to the others species, but leaves maintained smaller in both years since both average length and width remained smallest compared to the other species (Table S1).

Pairwise comparison revealed that between *A. rugosa* ‘After Eight’ and *A. rugosa*, five out of eight agronomic traits in both years presented significantly different values. Between *A. mexicana* and *A. foeniculum* there were no significant differences between average number of verticillaster and average flower number per verticillaster in either of the two years (Table 3).

Correlations between agronomic traits

Based on correlation coefficients between the eight agronomic traits studied, was observed that in both years existed significantly positive correlations between inflorescence length and the following variables: plant height and shoot number (Figure 3). Also, the correlation between leaf length and leaf width maintains significant in both years. However, some changes occurred between years. Although the correlation between flower number per verticillasters and verticillasters number per inflorescence was significantly positive in both years, the strength of the relationship increased in the second year. Similarly, the positive correlation between leaf number and flower number increases in the second year (Table S2).

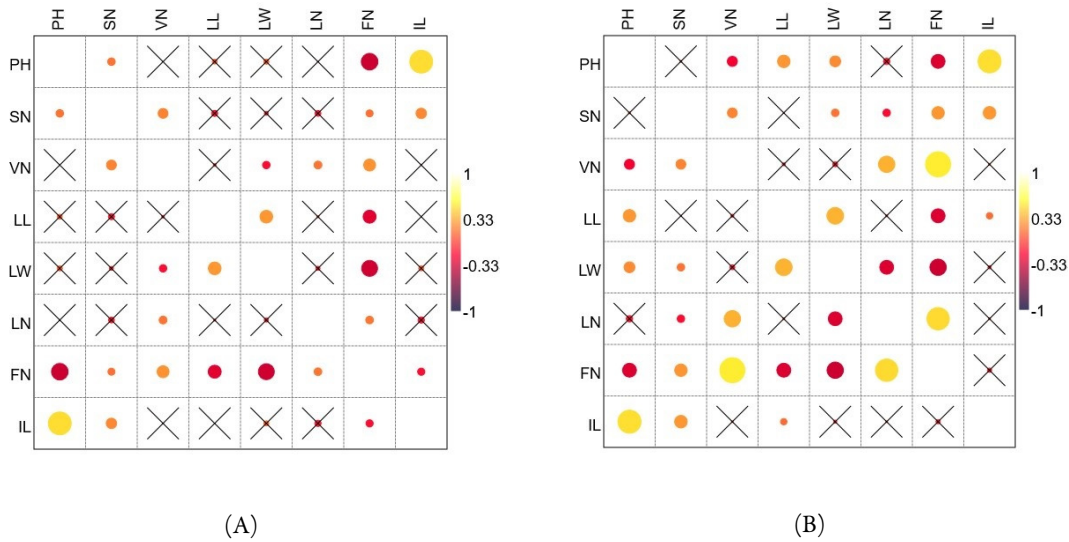


Figure 3. Correlograms presenting the relationship between morphometric traits of *Agastache* species: (A) 2019; (B) 2020
 Pearson correlations not significant at $p > 0.05$ = crossed “x”; size of bubble is proportional to the coefficient value.

Previsioning interrelationships between parameters

In 2020 the crop was considered acclimatized and it was attempted a previsioning of the possible plant architecture based on measured parameters. There were generated scenarios simulating the interdependence between agronomic traits and their variations. Thus, the model explores estimators of the relationship between predictor morphometric variables and the expected response. The interrelationships between the parameters of the studied species were analysed based on the AIC coefficient, being extracted the regression equations (Table 4).

According to the model, for *A. rugosa* ‘After Eight’, regarding the inflorescence’s length at 14.14 cm is added significantly 0.27 cm for each leaf that the plant develops, and 0.38 cm for each shoot. The calibration model was negatively set by the plants size and significantly positive increased by the flower number. In the case of *A. foeniculum* flower number, the basis established for calculating the was 24.79 flowers per verticillaster, to which was added a significantly 0.11 for each cm of the observed height, respectively 0.56 for each shoot developed. The width of the leaves decreases this parameter by 1.22 for every 1 cm. The inflorescences length has a high stability, the basic value being 7.28 cm, completed with 0.16 cm for each plant shoot (Table 4).

The height of *A. mexicana* plants is a trait than influences both flower number and inflorescence length. When analysing the flower number, the size was significantly influenced by plant height (0.28/1 cm), while the inflorescence length strongly reduces this parameter due to the lower number of verticillasters (-0.13/1 verticillasters).

Table 4. The model assessment (AIC) for *Agastache* flower number and inflorescence length and their interrelationships with the other analysed morphological parameters in the second vegetation year (2020)

Flower number	Indicator	Intercept	PH	SN	LL	LW	LN	VN	
<i>A. rugosa</i> ‘After Eight’	R ²	34.60							
	p value	<0.001							
<i>A. foeniculum</i>	R ²	24.79	0.11	0.56		-1.22			
	p value	0.001	0.041	0.065		0.112			
<i>A. mexicana</i>	R ²	12.80	0.28						
	p value	0.369	0.043						
<i>A. rugosa</i>	R ²	1.67	0.03	0.05	0.41				
	p value	0.461	0.138	0.029	0.013				
<i>A. scrophulariifolia</i>	R ²	-7.55		-0.43	2.31		0.41		
	p value	0.525		0.141	0.086		0.014		
All	R ²	21.52	-0.09	0.82	-1.05	-1.54	0.37	0.94	
	p value	5.93	0.041	0.14	0.34	0.474	0.06	0.171	
Inflorescence length	Indicator	Intercept	PH	SN	LL	LW	LN	VN	FN
<i>A. rugosa</i> ‘After Eight’	R ²	14.14	-0.11	0.38			0.27		-0.13
	p value	0.115	0.102	0.080			0.016		0.040
<i>A. foeniculum</i>	R ²	7.28		0.16					
	p value	<0.001		0.102					
<i>A. mexicana</i>	R ²	4.89	0.05					-0.13	
	p value	0.244	0.159					0.161	
<i>A. rugosa</i>	R ²	2.19		0.53		6.06	-0.35	0.40	
	p value	0.874		0.015		0.004	0.069	0.149	
<i>A. scrophulariifolia</i>	R ²	3.25					0.15		
	p value	0.179					0.0156		
All	R ²	-3.71	0.120	0.33		-0.720	0.07		-0.080
	p value	0.07	<0.001	<0.001		<0.001	<0.001		0.001

*Notes (legend): plant height (PH), shoot number (SN), leaf length (LL), LW (leaf width), leaf number (LN), verticillasters number (VN), flower number (FN).

Examination of coefficients for *A. rugosa* indicated a large number of parameters involved in determining the flower number and their length. The flower number has a low calculation base (1.67), but is significantly increased by 0.05 units/1 shoot, respectively 0.41 units/1 cm of leaf length. The inflorescences length is significantly higher due to the width of the leaves, 6.06 cm/1 cm wide, significantly supported by the

shoot number and the number of verticillasters. Instead, the pattern is negatively correlated by the leaf number (not significant), with a reduction in inflorescence length of 0.35/1 leaf.

A. scrophulariifolia had a high number of parameters influencing the flower number. Increase both in leaf length and leaf number being associated with increased flower number. Contrary, the number of shoots have a reducing effect for the inflorescence length. At an inflorescence length of 3.25 cm as base, is added 0.15 cm/1 developed leaf.

Projection of studied species based on the morphometric parameters

Projection based on the agronomic traits analysed, resulted in the clustering of species that enables visual identification of similarity or divergence among them (Figure 4). The first two components obtained explain over 70% of the variance observed in both years (Table S3). The cluster of data points corresponding to each species is packed closer together in the second year compared to the first, fact that might indicate to the stabilization of all species. The species *A. foeniculum* maintained a certain degree of overlapping with other species in both years, but notably with *A. rugosa*. In the first year, *A. foeniculum* also presented a certain overlapping with *A. scrophulariifolia* but this similarity did not maintain in the second year. The cultivar *A. rugosa* ‘After Eight’ clusters on plant height vector in both years, because it maintained the tallest during the experiment. The PCA analysis indicates that the analysed species expressed their phenotypic particularities during the studied interval, but differences become pronounced as plants become established and, evidenced by their clearer distinction in the second year.

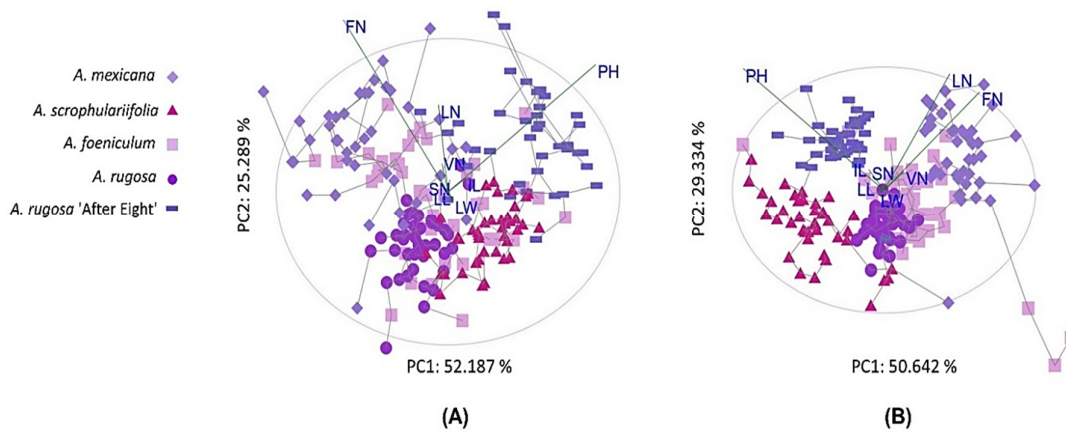


Figure 4. Principal component analysis of *Agastache* species displaying the least distance spanning tree, where vectors represent the eight agronomic traits studied in: (A) 2019, (B) 2020; ellipse 95% confidence interval; component values (PC1, PC2) indicate percentage of explained variance

Discussion

In the current context, the growers of medicinal crops are presented with the opportunity to extend their range of cultivated species, but they face difficulties at implementing the cultivation phase. These arise from the lack of informed knowledge to generate technological packages. Thus, agronomists must engage into defining suitability for cultivation and providing a consistent basis of knowledge and recommendations for local growers to enable them to overcome challenges related to extending their range of cultivated medicinal species.

Although the literature describes five stages of transition from wild harvesting to possible cultivation, the fifth stage called “cultivation phase” representing the stage when formal cultivation systems are designed

and instituted (Schippmann *et al.*, 2002) might benefit from a broadened and detailed consideration. This stage could be enlarged, to address also challenges related to species proposed for cultivation in other geographical areas, where their behaviour is known. Establishing best technological approaches, agronomic calendar of agricultural activities in relation with the behaviour and development of the species either new in cultivation or new to the area, requires a methodical approach. The stages of knowledge that could be considered when extending the cultivation of species from other geographical regions are summarized in Figure 5.

By applying adequate cultivation technology, the plant growth and development can be optimized to achieve biologic uniformity or high level of active principles (Muntean *et al.*, 2016; Schippmann *et al.*, 2002). In addition, cultivation can ensure a steady flow of raw material including from non-native species through a short-value chain (Schippmann *et al.*, 2002). Further selection and breeding efforts can give rise to highly productive genotypes (Wang *et al.*, 2020). Cultivation can also enable accurate planning of harvesting moment and processing of vegetal biomass (Vârban *et al.*, 2021).

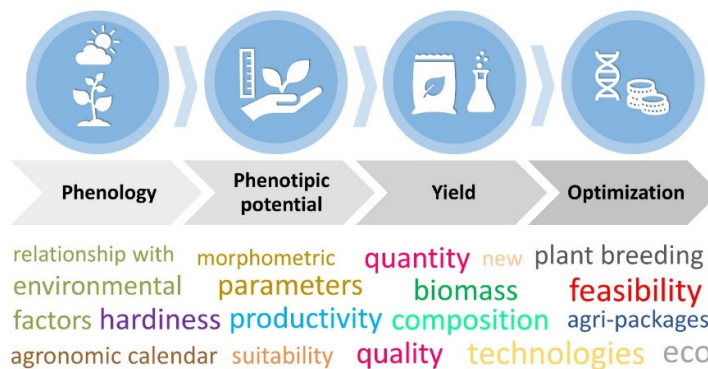


Figure 5. The interlinked stages within cultivation phase for species introduced/extended into cultivation in new areas

In the context of increasing demand of herbal remedies, safety and quality also come into forefront. Thus, at the European level, the Committee on Herbal Medicinal Products indicates in the “Guideline on Good Agricultural and Collection Practice” (GACP) that in the cultivation of medicinal plants the substances for growth and crop protection should be avoided or kept at the minimum while fertilizing agents should be applied sparingly (EMEA, 2006). In addition, World Health Organization announces that it is in the process of implementing the Traditional Medicine Strategy until 2023 (WHO, 20019). These guiding instruments provide principles and directions to be followed that supports a long-term and increasing contribution of medicinal plants to health and well-being.

Although there is an opportunity for growers to extend their range of cultivated species in the current context, the cultivation has to ensure a safe and natural biologic product. This springs up from increased demand of medicinal plants worldwide, that is fuelled by the association of their use with health promoting properties and a healthy lifestyle. Therefore, it is understandable why the final consumer generally prefers products derived from medicinal plants such as herbal medicines, teas, essential oils and extracts in cosmetics that have been grown as natural as possible. One of the most economically important botanic families for medicinal and aromatic purposes is Lamiaceae, that comprises some of the most widely used herbs, therefore the interest for species from this family, such as *Agastache* spp., is high (Hernandez-Leon *et al.*, 2021).

Based on this study, were determined the morphometric characteristics of *Agastache* sp. in local conditions. Overall, the studied species/cultivar displayed promising agronomic traits, and thus with promising potential to be grown and extended in cultivation, either for ornamental or for medicinal purposes. The average values for all parameters were higher in the second year, while the relationship between some traits tightens. The projection revealed a clearer distinction in second year. All these indicate to the acclimatization of the

studied species. Similarly, monitorization of acclimatization process of *A. foeniculum* conducted in Kremenets Botanical Garden from Ukraine between 2013 and 2015, reported also an increase in plant height from first (75.3 cm) to second year (135.7 cm) (Melnychuk and Rakhmetov, 2016). There is evidence that with age the presence of major compounds also changes. Content of rosmarinic acid, apigenin glucoside, chlorogenic acid were shown to be significantly differed between *A. rugosa* plants of different age (Bielecka *et al.*, 2019).

Plant height of *A. rugosa* in this study situates within the threshold of 60-120 cm mentioned in literature (Lord, 2003). In nine collections of *A. rugosa* from South Korea, grown under field conditions, average stem length was 88.86 cm, lateral branch number 23.93, leaf 9.80×6.88 cm, while the inflorescence length was 12.27 cm (Ok and Chae, 1998). By comparison, in our study *A. rugosa* presented taller plants in second year than reported by the authors cited, but the inflorescence and leaf size were smaller. Genotypes collected from seven regions of South Korea presented a plant height of 151.5 cm, leaf size of 8.4×6.1 cm, inflorescence length of 13.7 cm (Lee *et al.*, 1999). The parameters obtained in our study are lower for this species, however, one has to consider that East Asia is the native range, and therefore it could be expected that plants present superior values of morphometric parameters there. In a later study was showed that the application of calcium nitrate ensured increased average values of stem and inflorescence length of *A. rugosa*, as well as increased leaf length and width. Under different N application rates, the average shoot length was 75.2-81.5 cm, inflorescence length 10.2-11.1 cm, leaf length 8.3-8.9 cm, leaf width 5.6-6.2 cm. Study also showed that estragole content at blooming remained higher in leaves compared to inflorescence at all N fertilization levels tested or under different N forms (Ohk *et al.*, 2000), outlining the importance of leaves as source of bioactive compounds. Such results come to highlight the existing optimization possibilities by applying adequate technology.

A study conducted in Mexico on the domestication of *A. mexicana*, indicated that encouraged genotypes presented a plant height of 98.80 cm and inflorescence length of 19.73 cm while cultivated *A. mexicana* presented a vegetative height of 90.53 cm and inflorescence length of 12.15 cm (Carrillo-Galván *et al.*, 2020). According to the values obtained in our study, *A. mexicana* presented taller plants in second year than those registered by either encouraged or cultivated genotypes from Mexico, but average inflorescence length had smaller values. This could be attributed to the climate of Romania, that is colder, more humid, while solar radiation lower considering latitudinal position, and all these probably favoured the vegetative development.

A study conducted on some *Agastache* species grown under a hydroponic culture system for 4 weeks, indicated that *A. foeniculum* was outperformed regarding vegetative parameters by some *A. rugosa* cultivars. Thus, while *A. foeniculum* presented a stem length <30 cm in height and lower leaf number (<20), both *A. rugosa* 'Spike snow' and *A. rugosa* 'Spike blue' presented a stem over 30 height and higher leaf number (>20) after 4 weeks of cultivation in hydroponic system (Do *et al.*, 2020). This suggests that while through selection some cultivars are better suited for hydroponic cultivation, *A. foeniculum* exhibits less satisfactory results in such conditions. Another study conducted in pot conditions in greenhouse, reported for *A. foeniculum* an average plant height on 10th leaf of 118.11 cm for the diploid plants and of 85.98 cm for the tetraploids. Also, average inflorescence length was 5.19 cm in the diploid plants and 16.82 cm in the tetraploid ones (Talebi *et al.*, 2017). Genetic engineering and plant breeding certainly present some interesting possibilities to customize genotypes able to express maximized agronomic traits under either protected or field conditions and can represent a viable approach in optimizing the cultivation of these medicinal species. There is also emerging evidence suggesting that through adequate technology the morphometric traits of *A. foeniculum* can be enhanced. In this sense, a study has showed that foliar application of 2 g/L^{-1} urea significantly increased the leaf number and plant height of *A. foeniculum* (Jahani *et al.*, 2018).

A. foeniculum is the only species from this genus that has been considered for cultivation in local conditions before. A study from 2013 conducted on *A. foeniculum* that took place in Cluj County, indicated a yield of 3.05-3.83 t/ha dry weight. Out of the *herba* production obtained, highest proportion was represented by leaves and branches, together comprising 61.64% of the *herba* production, followed by inflorescence and main stem with a lower contribution (Duda *et al.*, 2013). Although in general the generative characters are to a less extent influenced by environmental conditions and age of the plant, in this study both the vegetative and

generative parameters registered an increasing trend from one year to the next. This might be explained by the relationships existing between vegetative and generative traits, considering that inflorescences are actually terminal ramifications of the stems.

Since in the current study were assessed the morphometric parameters in the absence of any interventions, the values reported can be considered the natural phenotypic potential of these species/cultivar in local conditions.

Conclusions

Between market opportunities and implementing diversification of medicinal crops, there is a missing link arising from challenges and difficulties faced by growers in up-scaling cultivation of novel or niche medicinal crops in their area. The successful cultivation of medicinal plants is conditioned by the phenotypic expression of given genotypes in certain environmental conditions. Precise knowledge of the phenotypic potential is a starting point for assessing feasibility of medicinal plants.

The present work tried to define the phenotypic potential in local conditions (Cluj-Napoca, Romania) for some medicinal species from genus *Agastache* that are not native to Europe in order to identify the favourability of their cultivation.

The eight agronomic parameters studied increased in values in the second year as the plants acclimatized and became established. In the second-year after planting in field, the average height was 109.8 cm. The shoot number almost doubled between first and second year, suggesting the productivity of these species. Average inflorescences length was 8.3 cm in the first year (2019) and 9.6 cm in the second year (2020). Among the species/cultivars studied, *A. rugosa* 'After Eight' was characterized by tall plants with longest inflorescence in both years and could be recommended also for ornamental purposes. At the opposite was situated *A. mexicana* that was characterized by shortest plants and shortest inflorescence in both years, but high number of verticillasters and flowers. *A. foeniculum* presented smallest leaves in both years, opposite to *A. scrophulariifolia* which presented largest leaves in both years. In both years existed significantly positive correlations between inflorescence length and the following variables: plant height and shoot number.

The PCA projection indicated that the analysed species expressed their phenotypic particularities during the studied interval, but differences were more pronounced as plants became established, evidenced by their clearer distinction in the second year.

Based on the present research were put in evidence some differences between studied species regarding the agronomic traits studied. These can be considered for targeted improvement in breeding programs of these species in order to achieve genotypes with balanced growth and development and acclimatized to local conditions. Despite differences, all species studied can be considered for widening the range of cultivated medicinal species.

Authors' Contributions

Conceptualization, RV and DIV; methodology, RV, DIV, AS, AO, VaS and VS; software, AO, IC and VS; validation, RV and VS; formal analysis, RV, DIV, AS, AO, VaS, VS; investigation, RV, DIV, AS, AO, VaS, VS; resources, RV, DIV and AS; data curation, RV; writing – original draft preparation, RV, AS, AO, VaS, IC and VS; writing – review and editing, RV, RVi, ŞG and SV; visualization IC and VS; supervision, RV, RVi, ŞG and SV. All authors have read and agreed to the published version of the manuscript. All authors read and approved the final manuscript.

Ethical approval (for researches involving animals or humans)

Not applicable.

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Conflict of Interests

The authors declare that there are no conflicts of interest related to this article.

References

- Ahmad KMS, Ahmad I (2019). Chapter 1 - Herbal medicine: current trends and future prospects. In: Ahmad Khan MS, Ahmad I, & Chattopadhyay D (Eds.). *New Look to Phytomedicine*. Academic Press, pp 3-13. <https://doi.org/10.1016/B978-0-12-814619-4.00001-X>
- Anand S, Deighton M, Livanos G, Morrison PD, Pang ECK, Mantri N (2019a). Antimicrobial activity of *Agastache* honey and characterization of its bioactive compounds in comparison with important commercial honeys. *Frontiers in Microbiology* 10:263. <https://doi.org/10.3389/fmicb.2019.00263>
- Anand S, Deighton M, Livanos G, Pang ECK, Mantri N (2019b). *Agastache* honey has superior antifungal activity in comparison with important commercial honeys. *Scientific Reports* 9(1):18197. <https://doi.org/10.1038/s41598-019-54679-w>
- Anand S, Pang E, Livanos G, Mantri N (2018). Characterization of physico-chemical properties and antioxidant capacities of bioactive honey produced from Australian grown *Agastache rugosa* and its correlation with colour and poly-phenol content. *Molecules* 23(1):108. <https://doi.org/10.3390/molecules23010108>
- Argyropoulos D (2019). EIP-AGRI Focus Group plant-based medicinal and cosmetic products. https://ec.europa.eu/eip/agriculture/sites/default/files/fg35_starting_paper_2019_en.pdf
- Bielecka M, Zielińska S, Pencakowski B, Stafiniak M, Ślusarczyk S, Prescha A, Matkowski A (2019). Age-related variation of polyphenol content and expression of phenylpropanoid biosynthetic genes in *Agastache rugosa*. *Industrial Crops Products* 141:111743. <https://doi.org/10.1016/j.indcrop.2019.111743>
- Cadar RL, Amuza A, Dumitras DE, Mihai M, Pocol CB (2021). Analysing clusters of consumers who use medicinal and aromatic plant products. *Sustainability* 13(15):8648. <https://doi.org/10.3390/su13158648>
- Carmona-Castro G, Estrada-Soto S, Arellano-García J, Arias-Duran L, Valencia-Díaz S, Perea-Arango I (2019). High accumulation of tilianin in *in-vitro* cultures of *Agastache mexicana* and its potential vasorelaxant action. *Molecular Biology Reports* 46(1):1107-1115. <https://doi.org/10.1007/s11033-018-4570-4>
- Carović-Stanko K, Petek M, Grdiša M, Pintar J, Bedeković D, Herak Ćustić M, Satovic Z (2016). Medicinal plants of the family Lamiaceae as functional foods – a review. *Czech Journal of Food Sciences* 34(5):377-390. <https://doi.org/10.17221/504/2015-CJFS>
- Carrillo-Galván G, Bye R, Eguiarte LE, Cristians S, Pérez-López P, Vergara-Silva F, Luna-Cavazos M (2020). Domestication of aromatic medicinal plants in Mexico: *Agastache* (Lamiaceae)-an ethnobotanical, morpho-physiological, and phytochemical analysis. *Journal of Ethnobiology and Ethnomedicine* 16(1):22. <https://doi.org/10.1186/s13002-020-00368-2>
- Choi S-K, Seo Y-N (1993). Studies on the germination physiology, growth and component analysis of *Agastache rugosa* KUNTZE. *Korean Journal of Plant Resources* 6(2):147-154.

- Climate Cluj (2021). Temperature, climate graph, Climate table for Cluj - Climate-Data.org. (n.d.). Retrieved 2021 October 23 from <https://en.climate-data.org/europe/romania/cluj-511/>
- Corrigan EE (2002). *Agastache scrophulariifolia* (Willd.) Kuntze. Purple giant hyssop: Conservation and research plant for New England. New England Wild Flower Society Framingham. MA. p. 1-22. Retrieved 2021 October 23 from <https://www.nativeplanttrust.org/documents/25/Agasthescrophulariifolia.pdf>
- Criveanu H (2001). Agrometeorology. Risoprint.
- Do JW, Noh SW, Bok GJ, Lee HJ, Lee JW, Park JS (2020). Selection of optimal varieties suitable for indoor cultivation considering the growth and functional content of *Agastache* species. Journal of Bio-Environment Control 29(2):202-208. <https://doi.org/10.12791/KSBEC.2020.29.2.202>
- Duda M, Matei CI, Vârban DI, Muntean S, Moldovan C (2013). The results of cultivating the species *Agastache foeniculum* (Pursh) Kuntze at Jucu, CJ. Bulletin of University of Agricultural Sciences and Veterinary Medicine Cluj-Napoca. Agriculture 70(1):214-217. <https://doi.org/10.15835/buasvmcn-agr:9787>
- EMA (2006). Guideline on good agricultural and collection practice (GACP) for starting materials of herbal origin (Guideline EMA/HMPC/246816/2005). European Medicines Agency.
- Fuentes-Granados R, Widrlechner M, Wilson L (1998). An overview of *Agastache* research. Journal of Herbs, Spices & Medicinal Plants 6:69-97. https://doi.org/10.1300/J044v06n01_09
- González-Ramírez AE, González-Trujano ME, Hernandez-Leon A, Valle-Dorado MG, Carballo-Villalobos A, Orozco-Suárez S, ... López-Muñoz F J (2021). Limonene from *Agastache mexicana* essential oil produces antinociceptive effects, gastrointestinal protection and improves experimental ulcerative colitis. Journal of Ethnopharmacology 280:114462. <https://doi.org/10.1016/j.jep.2021.114462>
- Hammer Ø (2021). Past 4 - The Past of the Future software (4.08) [Computer software]. Natural History Museum. <https://www.nhm.uio.no/english/research/infrastructure/past/index.html>
- Harrell Jr FE, Dupont C (2021). Hmisc: Harrell Miscellaneous (4.6-0) [Computer software]. <https://CRAN.R-project.org/package=Hmisc>
- Hernandez-Leon A, Moreno-Pérez GF, Martínez-Gordillo M, Aguirre-Hernández E, Valle-Dorado MG, Díaz-Reval MI, ... Pellicer F (2021). Lamiaceae in Mexican species, a great but scarcely explored source of secondary metabolites with potential pharmacological effects in pain relief. Molecules 26:7632. <https://doi.org/10.3390/molecules26247632>
- Hosseini B, Moharrami L (2014). The survey effect of BAP and TDZ on direct shoot regeneration from nodal explant of *Agastache foeniculum*. Journal of Crops Improvement 16(2):459-473. <https://doi.org/10.22059/jci.2014.53055>
- Hwang J M; Lee M-H, Lee J-H, Lee J-H (2021). *Agastache rugosa* extract and its bioactive compound tilianin suppress adipogenesis and lipogenesis on 3T3-L1 cells. Applied Science 11:7679. <https://doi.org/10.3390/app11167679>
- Index Seminum - Hortus Agro-Botanicus Napocensis (2021). AcademicPress.
- Ivanov I G, Vrancheva R Z, Petkova N T, Tumbarski Y, Dincheva I N, Badjakov I K (2019). Phytochemical compounds of anise hyssop (*Agastache foeniculum*) and antibacterial, antioxidant, and acetylcholinesterase inhibitory properties of its essential oil. Journal of Applied Pharmaceutical Science 9(02):072-078. <https://doi.org/10.7324/JAPS.2019.90210>
- Jahani R, Hassani A, Samadi A (2018). Effect of foliar application of urea, aspartic acid and glutamic acid on growth, physiological and biochemical characteristics of Anise hyssop (*Agastache foeniculum*). Applied Soil Research 5(2):950107. Retrieved 2021 October 23 from http://asr.urmia.ac.ir/article_20518.html?lang=en
- Jang TS, Moon HK, Hong SP (2015). Sex expression, population structure, and floral dimorphism in a gynodioecious herb, *Agastache rugosa* (Lamiaceae) in Korea. Flora - Morphology, Distribution, Functional Ecology of Plants 215:23-32. <https://doi.org/10.1016/j.flora.2015.06.004>
- Khorsandi O, Hassani A, Sefidkon F, Shirzad H, Khorsandi AR (2010). Effect of salinity (NaCl) on growth, yield, essential oil content and composition of *Agastache foeniculum* Kuntz. Iranian Journal of Medicinal and Aromatic Plants Research 26(3):438-451. <https://doi.org/10.22092/ijmapr.2010.6807>
- Kim SJ, Park JE, Bok GJ, Kanth BK, Lam VP, Park JS (2018). High electrical conductivity of nutrient solution and application of methyl jasmonate promote phenylpropanoid production in hydroponically grown *Agastache rugosa*. Horticultural Science and Technology 36(6):841-852. <https://doi.org/10.12972/kjhst.20180082>
- Kovalenko NA, Supichenko GN, Leontiev VN, Shutova AG (2019). Composition of essential oil of plants some species of the genus *Agastache* L. Introduced in Belarus. Proceedings of the National Academy of Sciences of Belarus, Biological Series 64(2):147-155. <https://doi.org/10.29235/1029-8940-2019-64-2-147-155>

- Kuhn M, Wing J, Weston S, Williams A, Keefer C, Engelhardt A ... Hunt T (2021). caret: classification and regression training (6.0-90) [Computer software]. <https://CRAN.R-project.org/package=caret>
- Lam VP, Kim SJ, Bok GJ, Lee JW, Park JS (2020a). The effects of root temperature on growth, physiology, and accumulation of bioactive compounds of *Agastache rugosa*. Agriculture 10(5):162. <https://doi.org/10.3390/agriculture10050162>
- Lam VP, Kim SJ, Park JS (2020b). Optimizing the electrical conductivity of a nutrient solution for plant growth and bioactive compounds of *Agastache rugosa* in a plant factory. Agronomy 10(1):76. <https://doi.org/10.3390/agronomy10010076>
- Lee S-W, Kim J-B, Kim K-S, Kim M-S (1999). Changes of growth characteristics, rosmarinic acid and essential oil contents according to harvest time in *Agastache rugosa* O. Kuntze. Korean Journal of Medicinal Crop Science 7(2):83-88. Retrieved 2021 October 23 from <https://www.koreascience.or.kr/article/JAKO199903042330184.page>
- Li FS, Weng JK (2017). Demystifying traditional herbal medicine with modern approach. Nature Plants 3(8):1-7. <https://doi.org/10.1038/nplants.2017.109>
- Lint H, Epling C (1945). A revision of *Agastache*. The American Midland Naturalist 33(1):207-230. <https://doi.org/10.2307/2421328>
- Lord T (2003). Flora: The Gardener's Bible. Cassell - Weidenfeld & Nicolson.
- Marchioni I, Dimita R, Gioè G, Pistelli L, Ruffoni B, Pistelli L, Najar B (2021). The effects of post-harvest treatments on the quality of *Agastache aurantiaca* edible flowers. Horticulturae 7(4):83. <https://doi.org/10.3390/horticulturae7040083>
- Melnychuk OA, Rakhmetov DB (2016). The peculiarities of growth and development of *Lophanthus anisatus* Adans. Plants introducing in the conditions of Kremets Botanical Garden. Plant Introduction 72:39-44. <https://doi.org/10.5281/zenodo.2457491>
- Mendiburu F (2020). Agricolae: Statistical procedures for agricultural research. R package version 1.3-3. <https://CRAN.R-project.org/package=agricolae>
- Mohamed YA, Naser A (2012). Assessment of nitrate and nitrite contamination in herbal tea products. Journal of Medicinal Plants Research 6(19):3555-3560. <https://doi.org/10.5897/JMPR12.013>
- Muntean LS, Tămaș M, Muntean S, Muntean L, Duda MM, Vârban DI, Florian S (2016). Tratat de plante medicinale cultivate și spontane (II). Risoprint.
- Najafi F, Kavooosi G, Siahbalaeei R, Kariminia A (2022). Anti-oxidative and anti-hyperglycemic properties of *Agastache foeniculum* essential oil and oily fraction in hyperglycemia-stimulated and lipopolysaccharide-stimulated macrophage cells: *In vitro* and *in silico* studies. Journal of Ethnopharmacology 284:114814. <https://doi.org/10.1016/j.jep.2021.114814>
- Najar B, Marchioni I, Ruffoni B, Copetta A, Pistelli L, Pistelli L (2019). Volatilomic analysis of four edible flowers from *Agastache* genus. Molecules 24(24):4480. <https://doi.org/10.3390/molecules24244480>
- Nchu F, Matanzima Y, Laubscher CP (2017). Prospects of N fertilization in medicinal plants cultivation. In: Nitrogen in Agriculture. IntechOpen. <https://doi.org/10.5772/intechopen.68165>
- Ohk H-C, Song J-S, Chae Y-A (2000). Effect of forms and levels of nitrogen fertilizer on plant growth and essential oil content of *Agastache rugosa*. Korean Journal of Crop Science 45(2):128-133.
- Ok H-C, Chae Y-A (1998). Characteristics of seed and plant growth in local collections of *Agastache rugosa*. Korean Journal of Crop Science 43(4):269-272. Retrieved 2021 October 23 from <https://www.koreascience.or.kr/article/JAKO199811922913335.pdf>
- Oksanen J, Blanchet FG, Friendly M, Kindt R, Legendre P, McGlenn D ... Wagner H (2020). vegan: community ecology package (2.5-7) [Computer software]. <https://CRAN.R-project.org/package=vegan>
- Özcan MM, Akbulut M (2008). Estimation of minerals, nitrate and nitrite contents of medicinal and aromatic plants used as spices, condiments and herbal tea. Food Chemistry 106(2):852-858. <https://doi.org/10.1016/j.foodchem.2007.06.045>
- Palma-Tenango M, Sánchez-Fernández RE, Soto-Hernández M (2021). A systematic approach to *Agastache mexicana* research: biology, agronomy, phytochemistry, and bioactivity. Molecules 26(12):3751. <https://doi.org/10.3390/molecules26123751>
- Polivanova OB, Cherednichenko MY (2017). Introduction to in vitro Culture and Micropropagation of two *Agastache* species—*Agastache urticifolia* and *Agastache scrophulariifolia*. 100. Retrieved 2021 October 23 from <https://elib.bsu.by/bitstream/123456789/179997/1/p135.pdf>

- Quattrocchi U (2016). CRC world dictionary of medicinal and poisonous plants: common names, scientific names, eponyms, synonyms, and etymology (5 Volume Set). CRC Press.
- R: The R Project for Statistical Computing. (n.d.). Retrieved 2021 December 4 from <https://www.r-project.org/>
- Ripley B, Venables B, Bates DM, Hornik K, Gebhardt A, Firth D (2021). MASS: Support Functions and Datasets for Venables and Ripley's MASS (7.3-54) [Computer software]. <https://CRAN.R-project.org/package=MASS>
- Robinson D *et al.*, (2021). broom: Convert Statistical Objects into Tidy Tibbles (0.7.10) [Computer software]. <https://CRAN.R-project.org/package=broomRStudio>. Open source & professional software for data science teams. (n.d.). Retrieved 2021 December 4 from <https://rstudio.com><https://www.rstudio.com/>
- Rudik GO (2016). Morphological structure of inflorescences of *Agastache breviflora* (A. Gray) Epling, *A. rugosa* (Fisch. & amp; C.A. Mey.) Kuntze, and *A. rupestris* (Greene) Standl. (Family Lamiaceae) *ex situ*. Modern Phytomorphology 10:81-86.
- Schippmann U, Leaman D, Cunningham AB (2002). Impact of cultivation and gathering of medicinal plants on biodiversity: global trends and issues. FAO.
- Schippmann U, Leaman D, Cunningham AB (2006). A comparison of cultivation and wild collection of medicinal and aromatic plants under sustainability aspects. In: Bogers RJ, Craker LE, Lange D (Eds). Medicinal and Aromatic Plants 17:75-95. Springer Netherlands. https://doi.org/10.1007/1-4020-5449-1_6
- Sheahan CM (2012). Fact sheet for purple giant hyssop (*Agastache scrophulariifolia*). USDA-Natural Resources Conservation Service. Retrieved 2021 October 23 from https://plants.usda.gov/DocumentLibrary/factsheet/pdf/fs_agsc.pdf
- Talebi SF, Saharkhiz MJ, Kermani MJ, Sharafi Y, Raouf FF (2017). Effect of different antimetabolic agents on polyploid induction of anise hyssop (*Agastache foeniculum* L.). Caryologia 70(2):184-193. <https://doi.org/10.1080/00087114.2017.1318502>
- Vârban R, Ona A, Stoie A, Vârban D, Crişan I (2021). Phenological assessment for agronomic suitability of some *Agastache* species based on standardized BBCH Scale. Agronomy 11(11):2280. <https://doi.org/10.3390/agronomy11112280>
- Wang W, Xu J, Fang H, Li Z, Li M (2020). Advances and challenges in medicinal plant breeding. Plant Science 298:110573. <https://doi.org/10.1016/j.plantsci.2020.110573>
- WFO (2021). *Agastache* J. Clayton *ex* Gronov. Retrieved 2021 September 12 from <http://www.worldfloraonline.org/taxon/wfo-4000000903>
- WHO (2019) WHO Global Report on Traditional and Complementary Medicine; Geneva, Switzerland.
- Yeo HJ, Park CH, Park YE, Hyeon H, Kim JK, Lee SY, Park SU (2021). Metabolic profiling and antioxidant activity during flower development in *Agastache rugosa*. Physiology and Molecular Biology of Plants 27(3):445-455. <https://doi.org/10.1007/s12298-021-00945-z>
- Zielińska S, Matkowski A (2014). Phytochemistry and bioactivity of aromatic and medicinal plants from the genus *Agastache* (Lamiaceae). Phytochemistry Reviews 13(2):391-416. <https://doi.org/10.1007/s11101-014-9349-1>



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